

# Genetic bases of early root growth and rhizosheath formation in pearl millet (*Pennisetum glaucum* L.)

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microorganismes associés aux Stress Environnementaux (LAPSE),  
Dakar, Senegal



AfricaRice



Hangzhou, November 2018

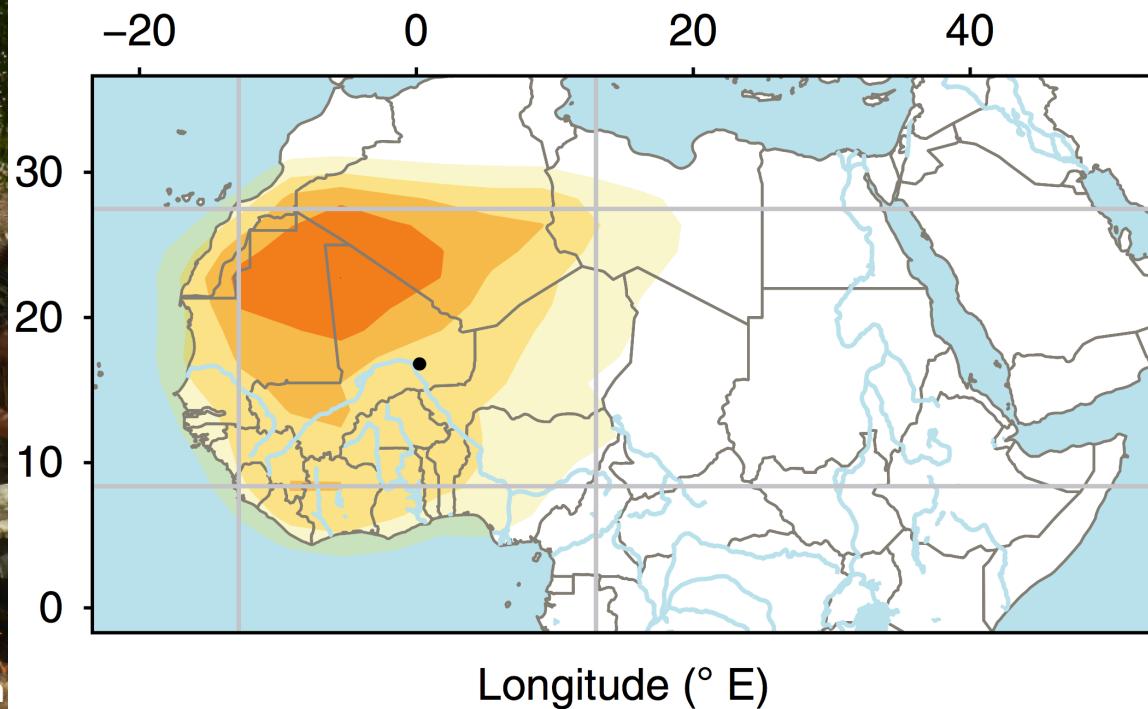


# Pearl millet

*Pennisetum glaucum*



Photos: C.T. Hash



Burgarella *et al*, Nat. Ecol. Evol. 2018



Hangzhou, November 2018

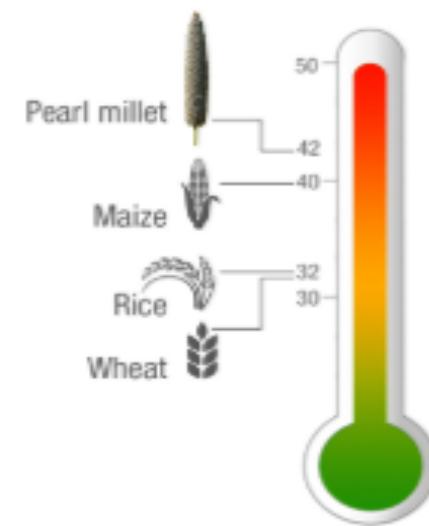
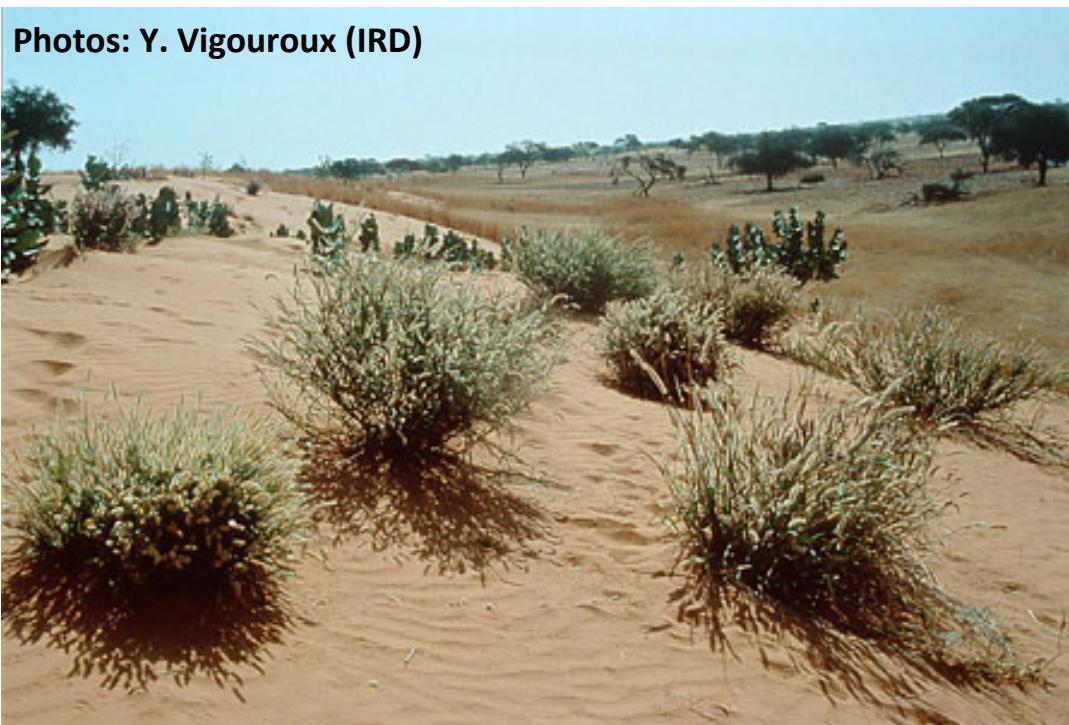


# Pearl millet

*One of the hardiest cereals*



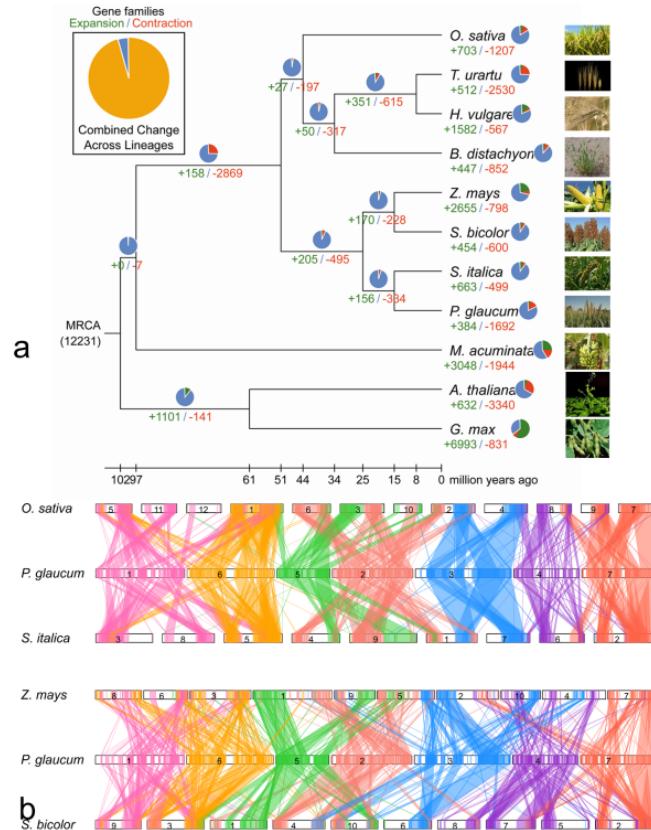
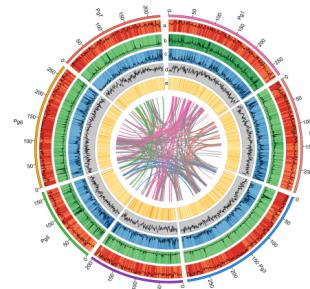
Photos: Y. Vigouroux (IRD)



Ref: ICRISAT Newsletter (<http://www.icrisat.org/>)



# Pearl millet Genome



Varshney et al, Nat. Biotech. 2017

- Consortium ICRISAT/BGI/IRD
- Genome size: 1.79 GB
- 38 579 predicted genes
- High GC (47.9%) and TE (80%) content
- Expansion genes families for cutin/suberin and ABC transporters
- 994 lines fully resequenced

Hangzhou, November 2018



# Pearl millet

*Main limiting factors*



- **Abiotic stresses**
  - Low soil fertility (low P availability)
  - Drought
  - Heat stress
- **Biotic stresses**
  - Striga
  - Fungal pathogens
  - Insects & nematodes
  - Birds



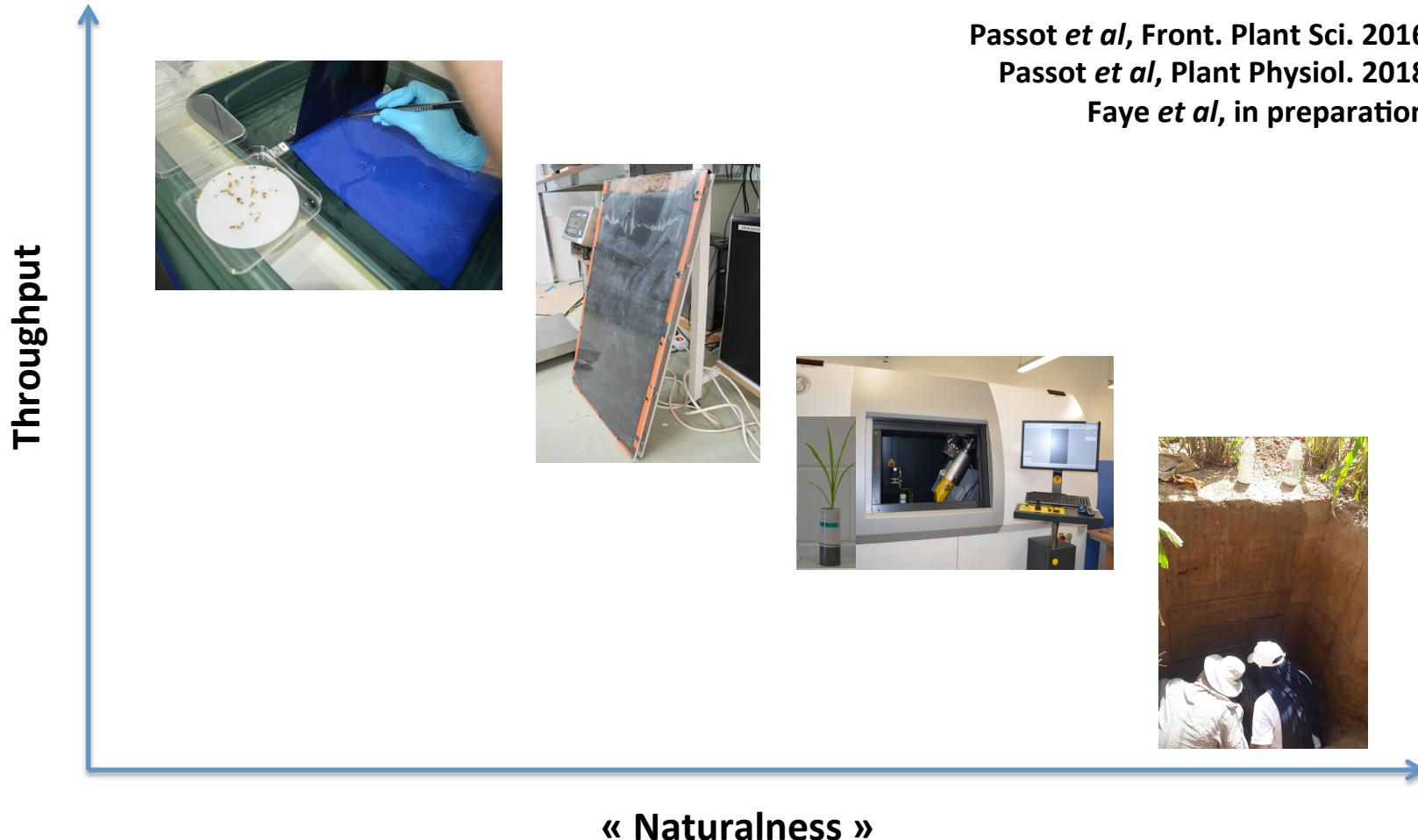
# Objectives

**Identify root traits that contribute to pearl millet adaptation to low water and low nutrients conditions**

- **Describe/quantify root trait**
  - *root architecture and anatomy*
  - *interactions with beneficial microbes*
- **Identify desirable root traits**
- **Identify genes controlling desirable root traits**



# Root phenotyping



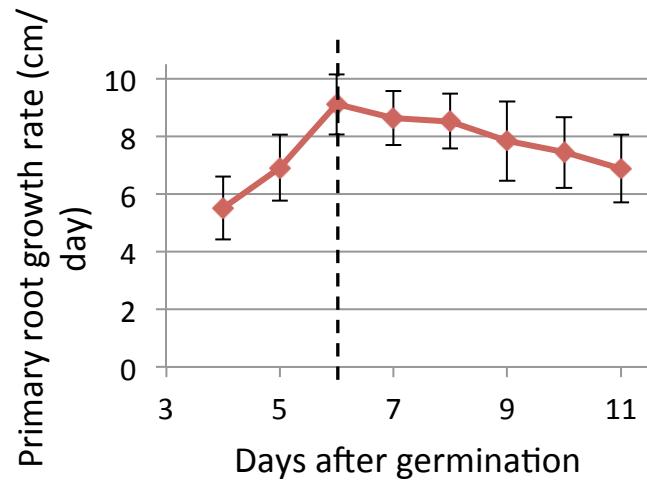
Hangzhou, November 2018



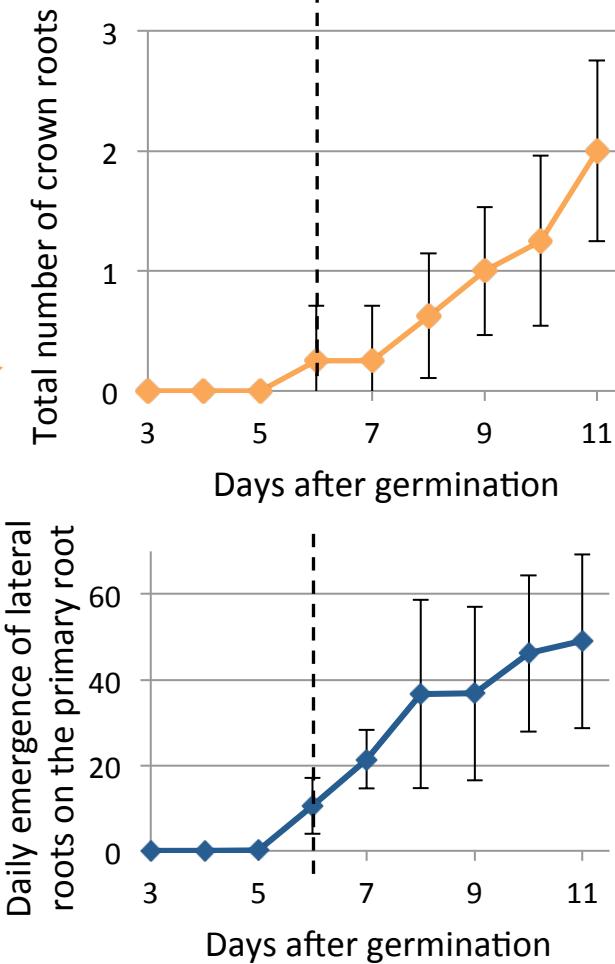
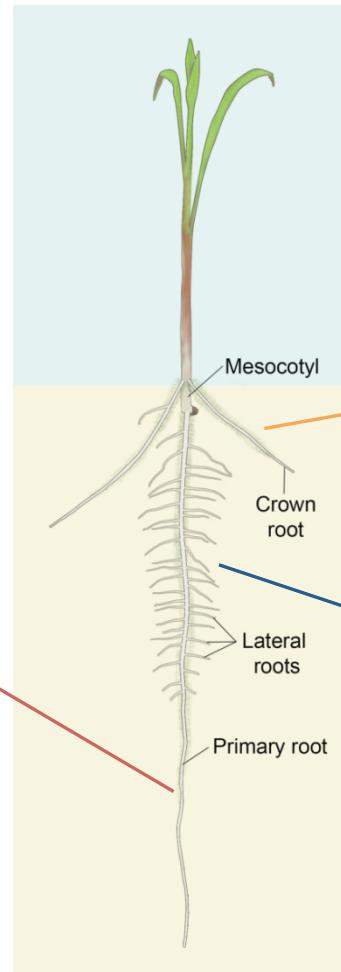
# Early developmental dynamics of pearl millet root system



S. Passot



Carbon reallocation to newly born root axes at 6 days



Passot *et al*, Front. Plant Sci. 2016

Passot *et al*, Plant physiol. 2018

Hangzhou, November 2018



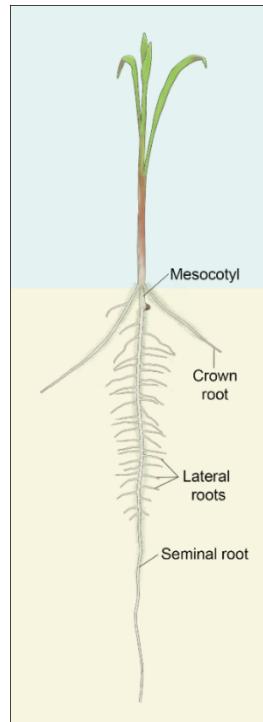
# Primary root growth

## *High-throughput phenotyping*



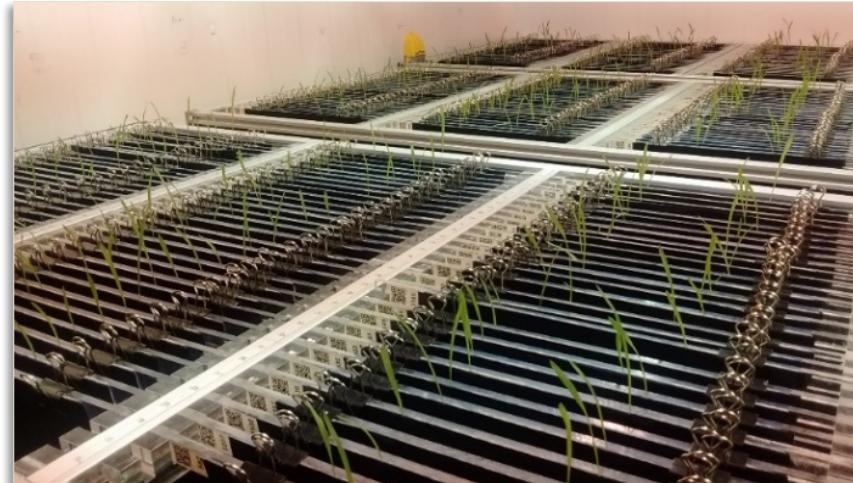
S. Passot

Pearl millet early  
root system



Very fast growth  
of seminal root

Hydroponic paper-based phenotyping platform



Hangzhou, November 2018



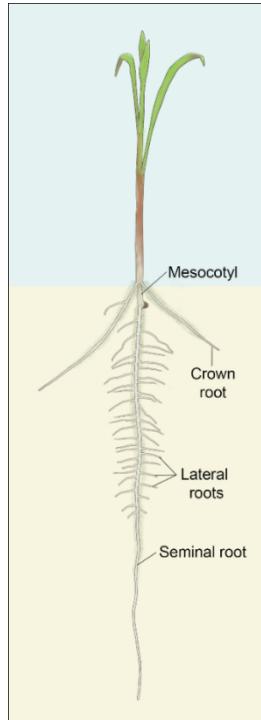
# Primary root growth

## *Genetic diversity*



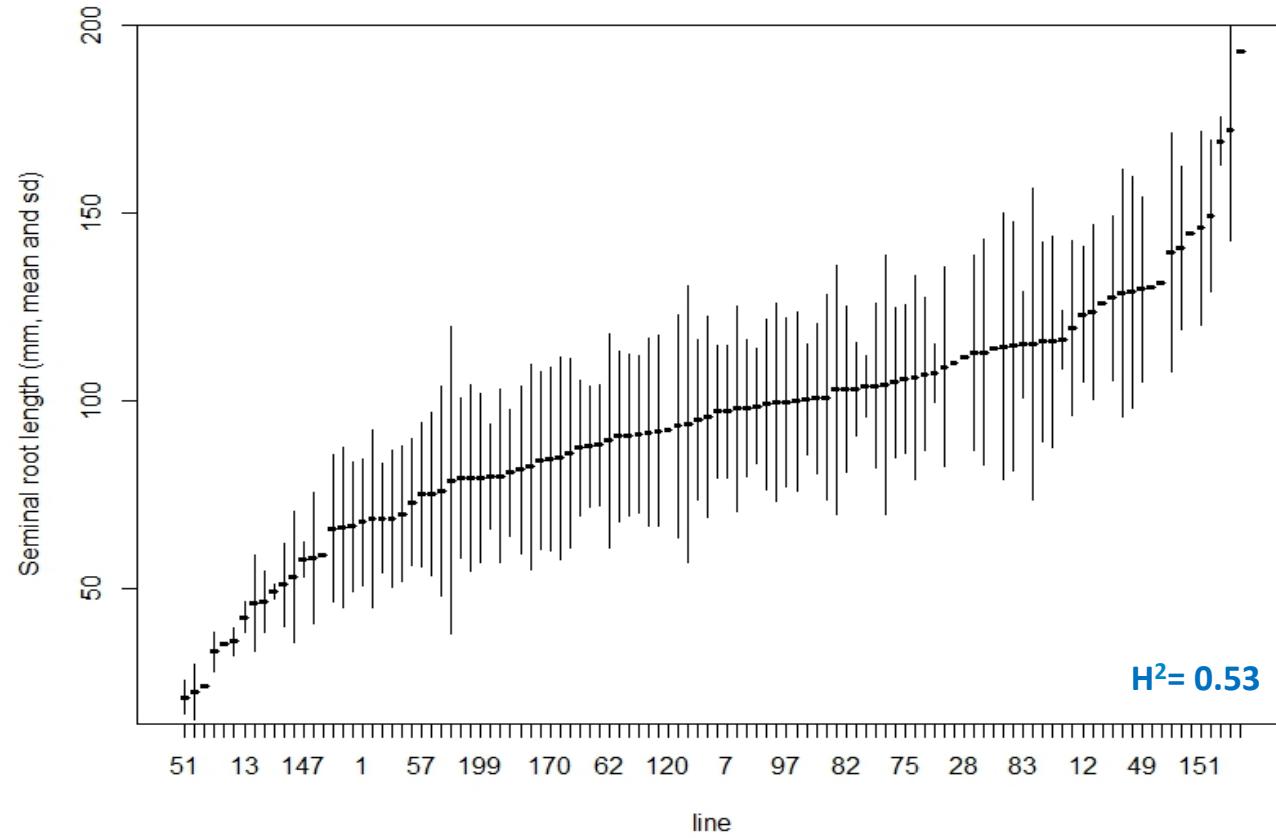
S. Passot

Pearl millet early  
root system



Very fast growth  
of seminal root

Seminal root length in a panel of 108 inbred lines 6 days after germination

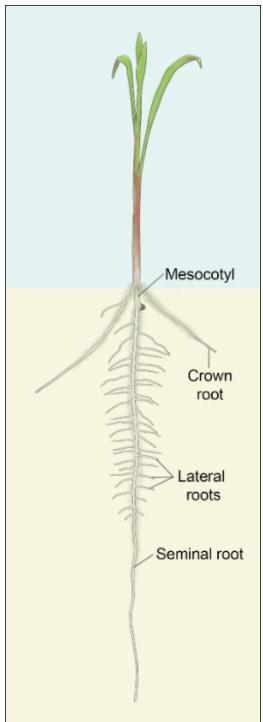




# Primary root growth

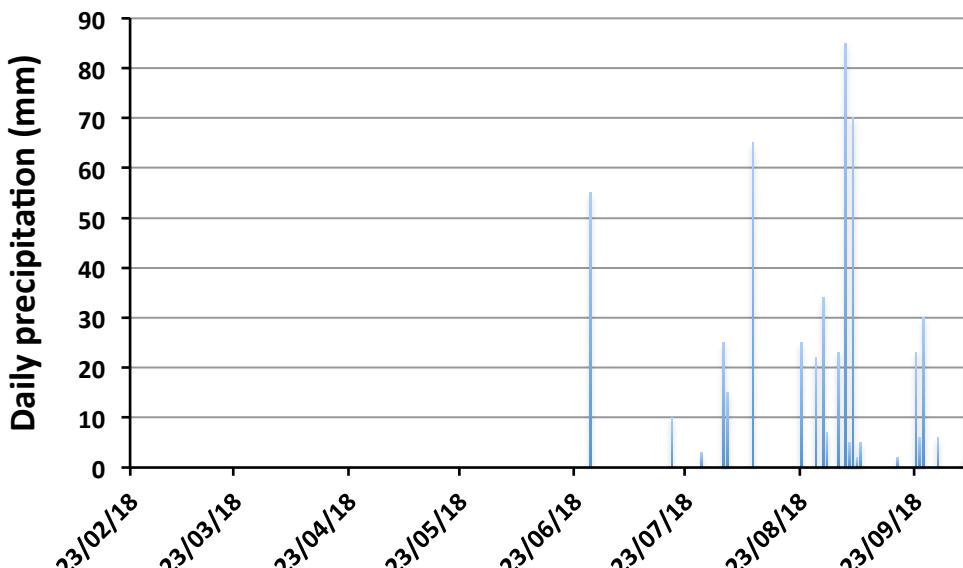
## *An adaptative trait?*

Pearl millet early  
root system



Very fast growth  
of seminal root

Precipitation pattern in Sob (Senegal) - 2018



Source: OPSE Niakhar



# Primary root growth

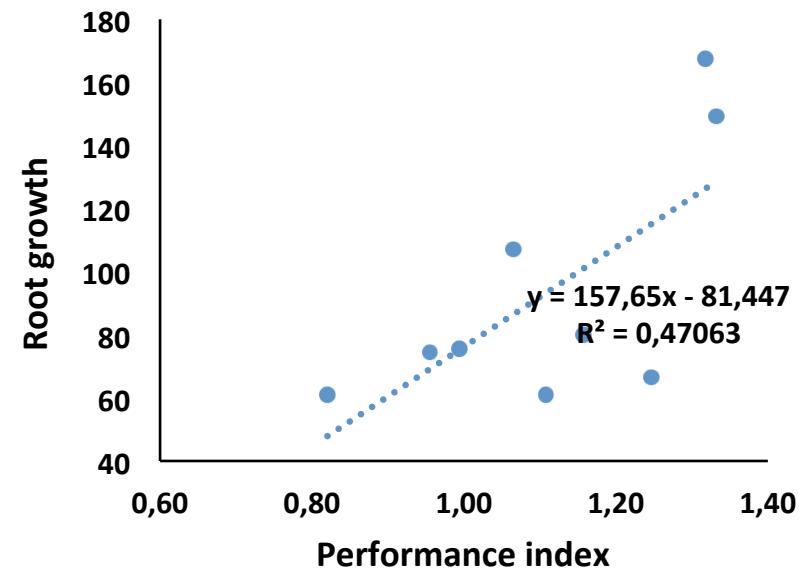
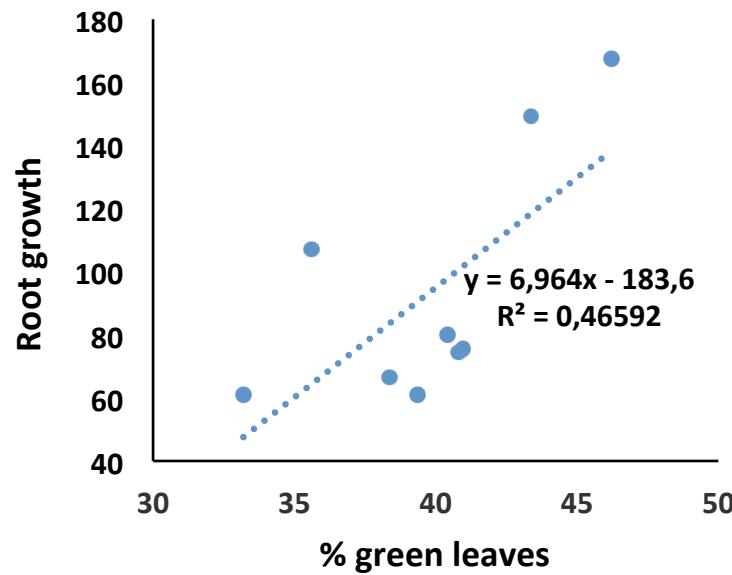
## *Field trials results*



B. Sine



A. Grondin





# Primary root growth

## *Field trials results*

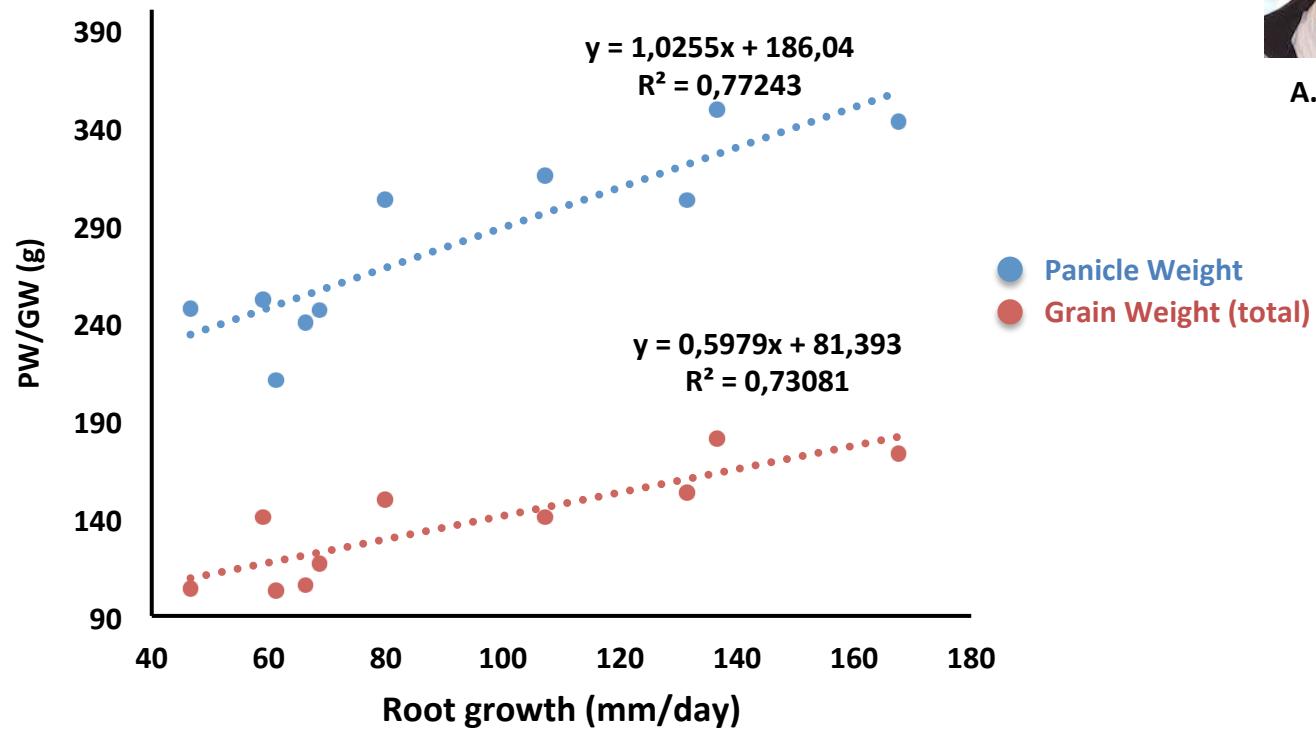


B. Sine



A. Grondin

Field trials (+/- P, WW/STR1/STR2)



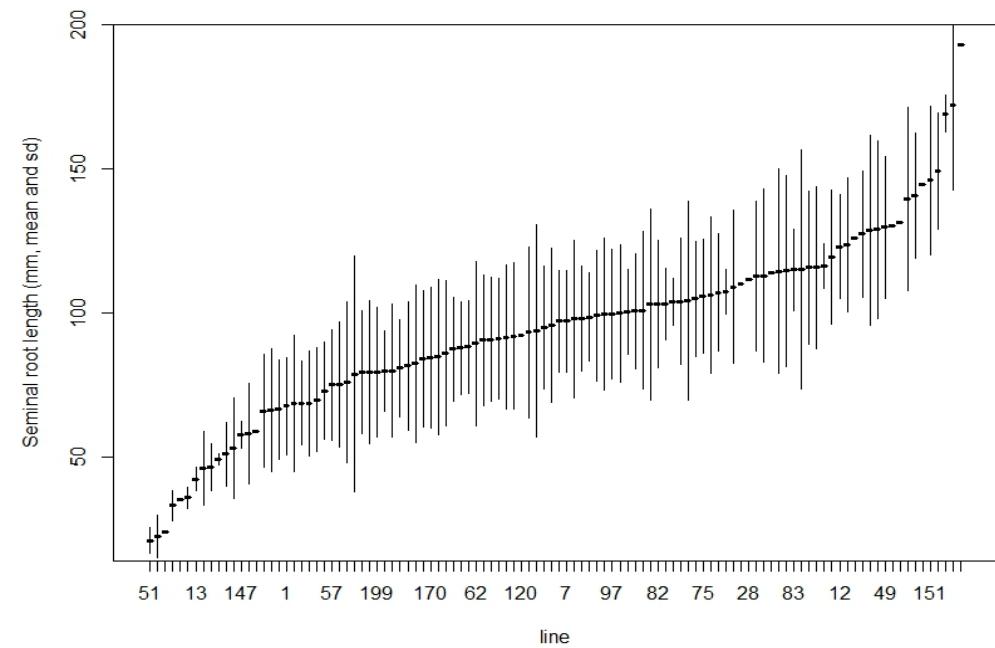
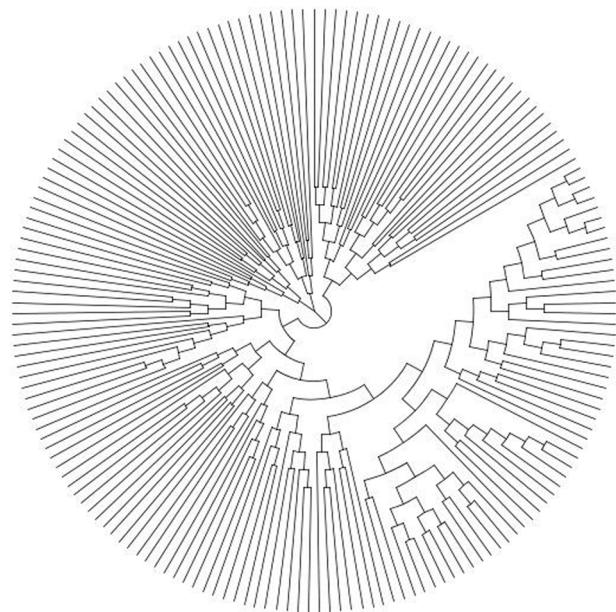


# Primary root growth GWAS



S. Passot

392 493 SNPs identified by GBS

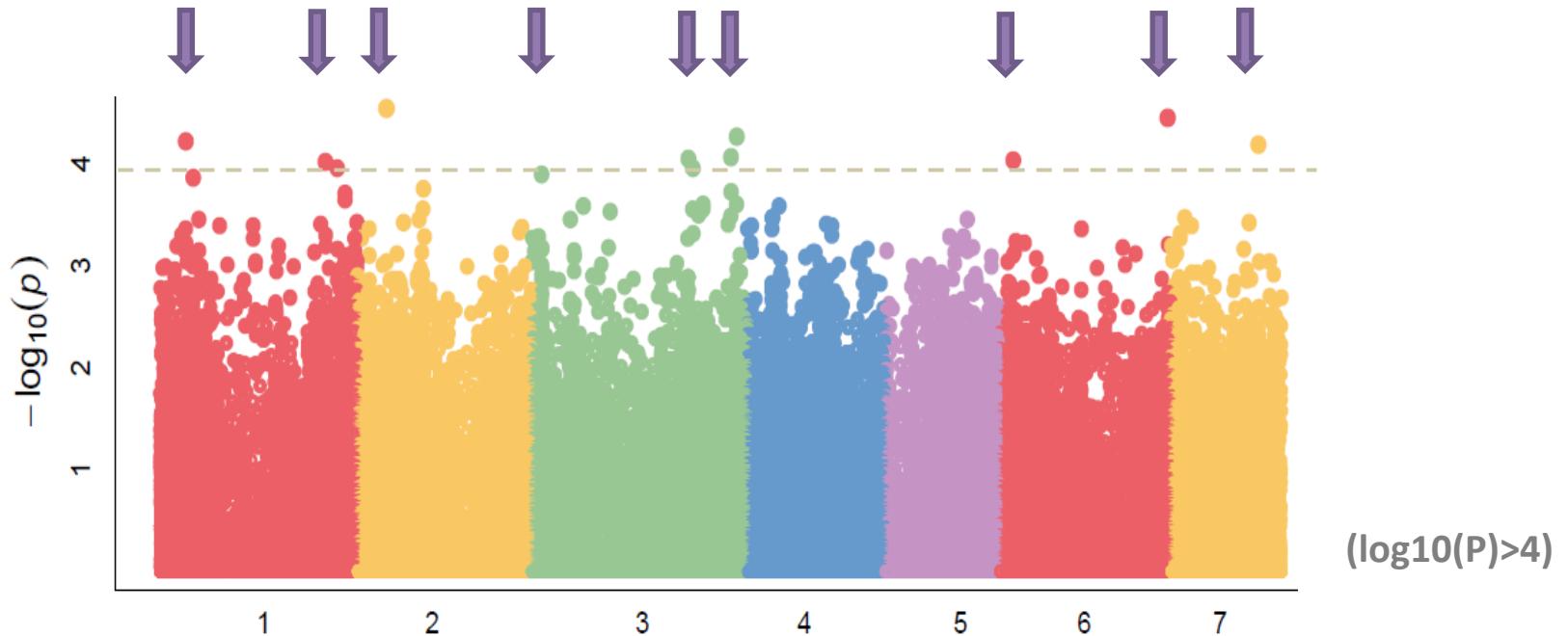




# Primary root growth GWAS

M. Debieu

9 potential QTLs controlling primary root length

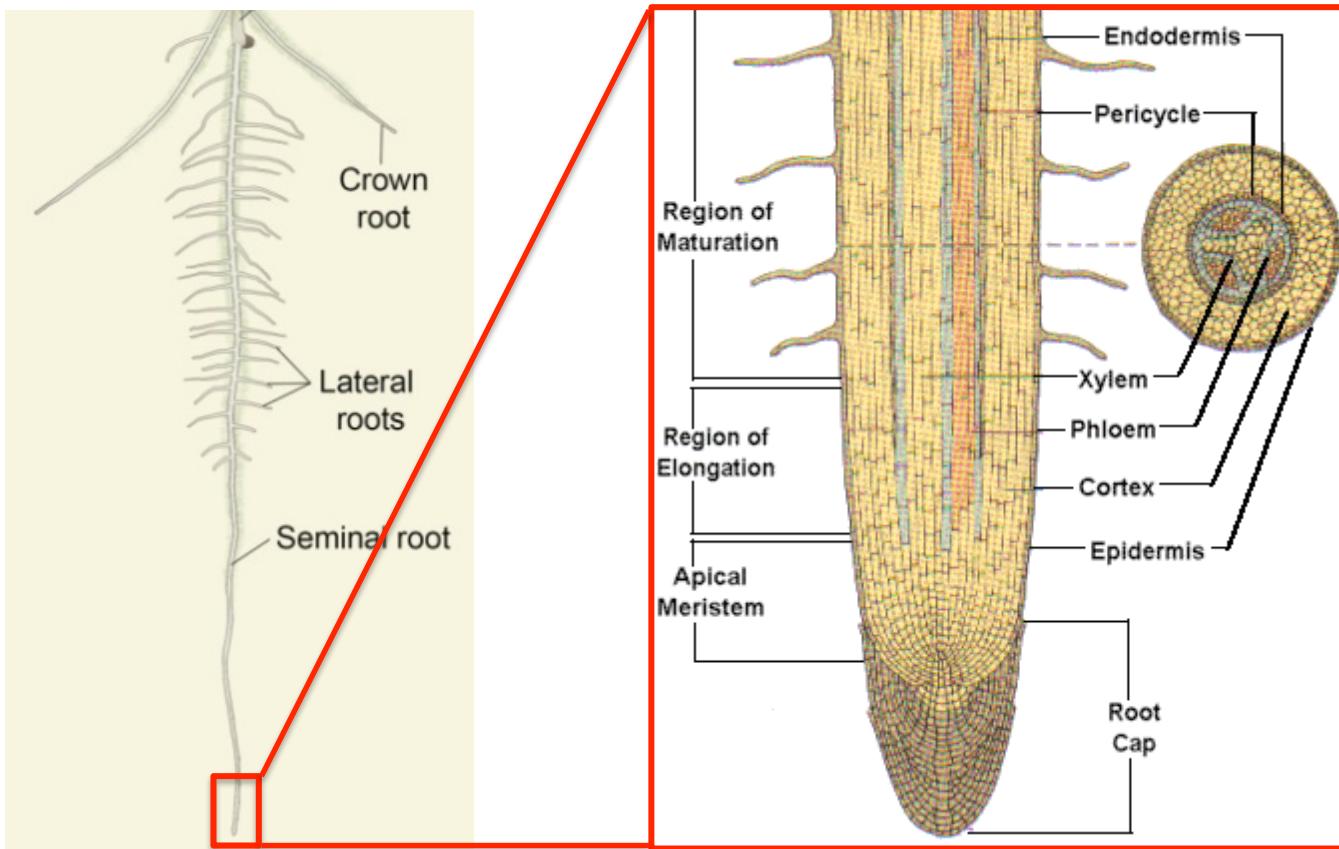


~ 150 genes in QTL regions (500 kb interval around most significant SNP markers)



# Primary root growth

## *Transcriptomics*



From <https://www.tes.com/lessons/nHBDO6YooXG09A/plant-anatomy-physiology>



# Primary root growth

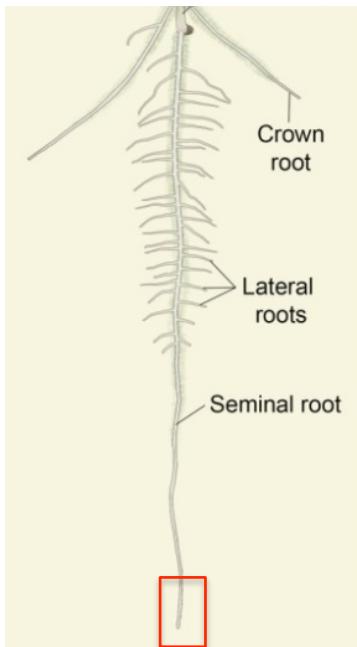
## *Transcriptomics*



M. Debieu



A. Grondin



RNAseq

Low  
growth  
IL 249

VS

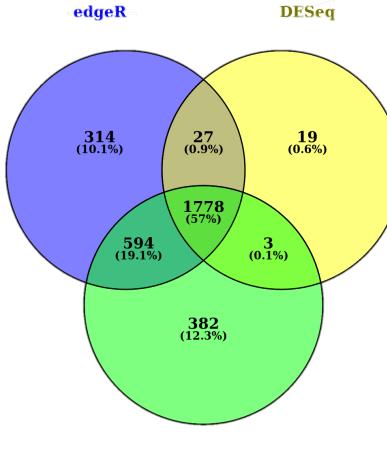
Montpellier  
**MGX**  
GenomiX

High  
growth  
IL 337

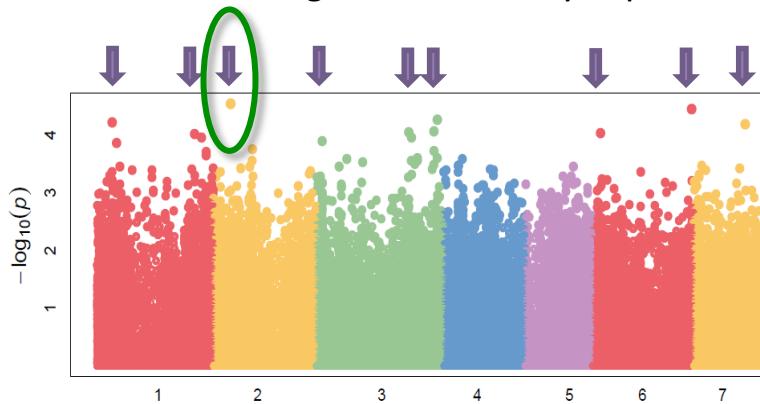
- 3 replicates
- single read 50 nt (Illumina HiSeq 2500)
- **1/3 reads not mapped to predicted CDS**
- 3 statistical tests (EdgeR, DESeq & DESeq2)



# Genetic bases root growth



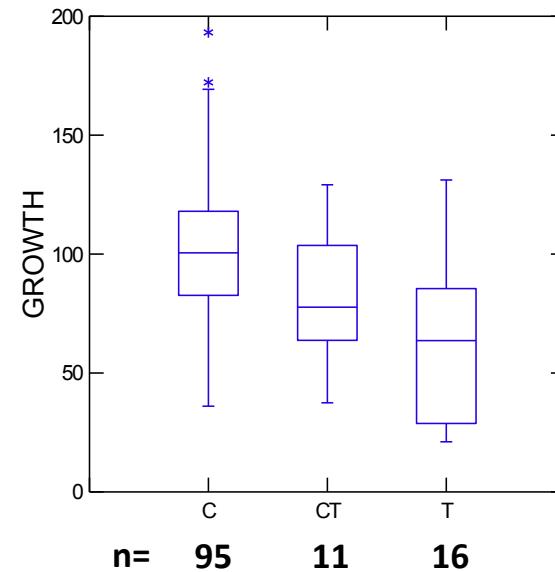
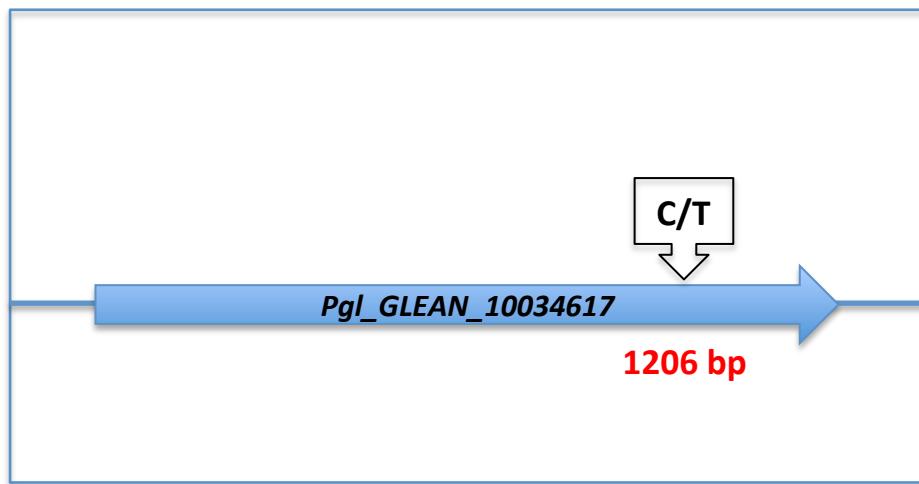
RNAseq data from root tips of two contrasting lines revealed 1778 genes differently expressed



- ~ 100 expressed genes in QTL regions
- 3 SNPs located within genes with predicted functions for root growth



# Genetic bases root growth



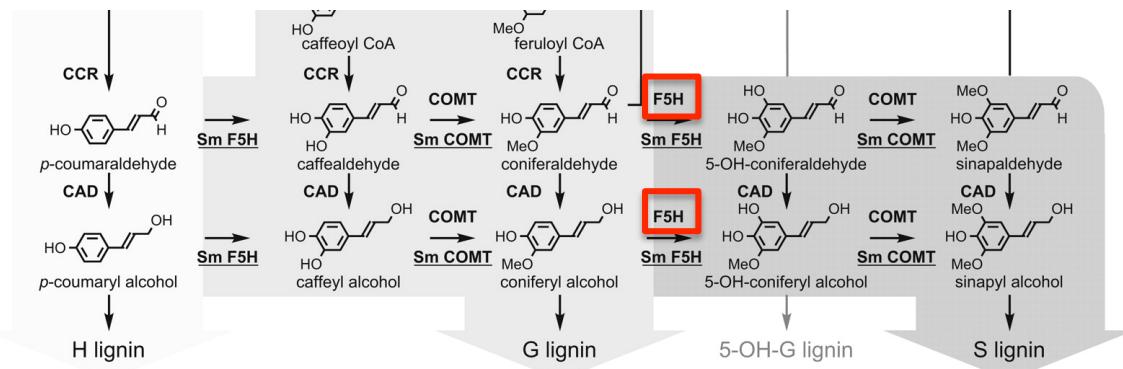


# Genetic bases root growth

## Progressive Inhibition by Water Deficit of Cell Wall Extensibility and Growth along the Elongation Zone of Maize Roots Is Related to Increased Lignin Metabolism and Progressive Stelar Accumulation of Wall Phenolics<sup>1</sup>

Ling Fan, Raphael Linker, Shimon Gepstein, Eiichi Tanimoto, Ryoichi Yamamoto, and Peter M. Neumann\*

Plant Physiology Laboratory, Department of Environmental, Water, and Agricultural Engineering, Faculty of Civil and Environmental Engineering (L.F., R.L., P.M.N.) and Faculty of Biology (S.G.), Technion-Israel Institute of Technology, Haifa 32000, Israel; Plant Physiology Laboratory, Department of Information and Biological Sciences, Graduate School of Natural Sciences, Nagoya City University, Nagoya 467-8501, Japan (E.T.); and Biology and Chemistry Laboratory, Tezukayama University, Nara 631-8585, Japan (R.Y.).

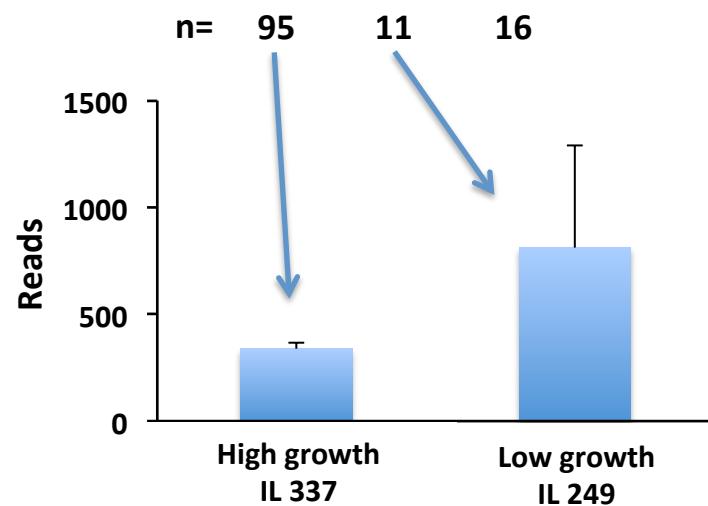
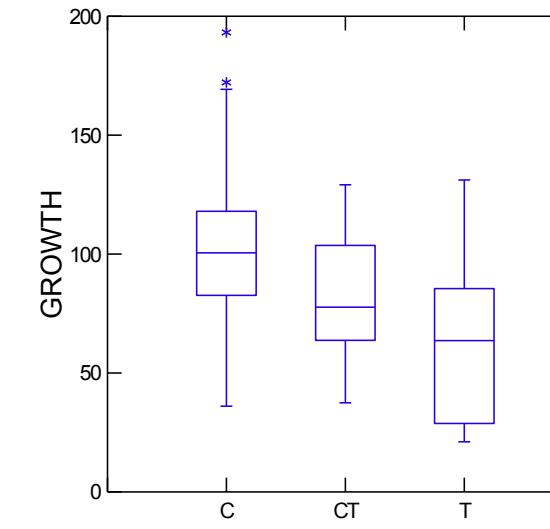
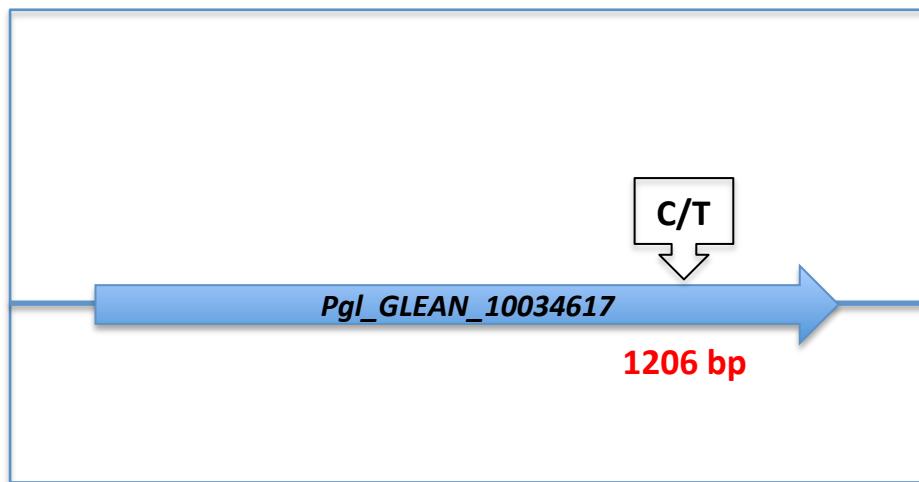


Jing-Ke Weng et al. *Plant Cell* 2011;23:2708-2724

Hangzhou, November 2018



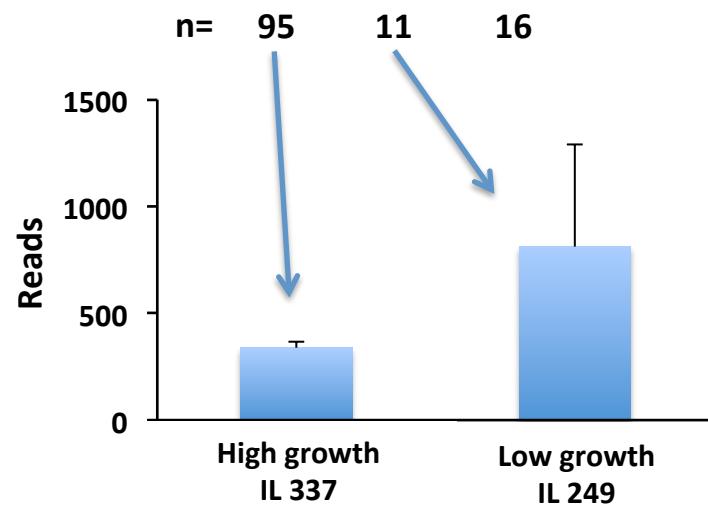
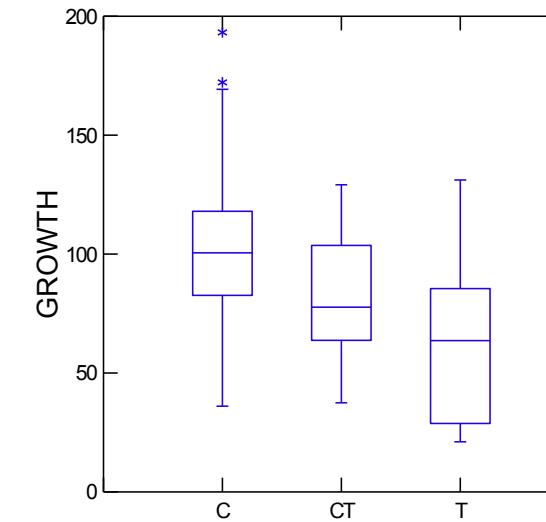
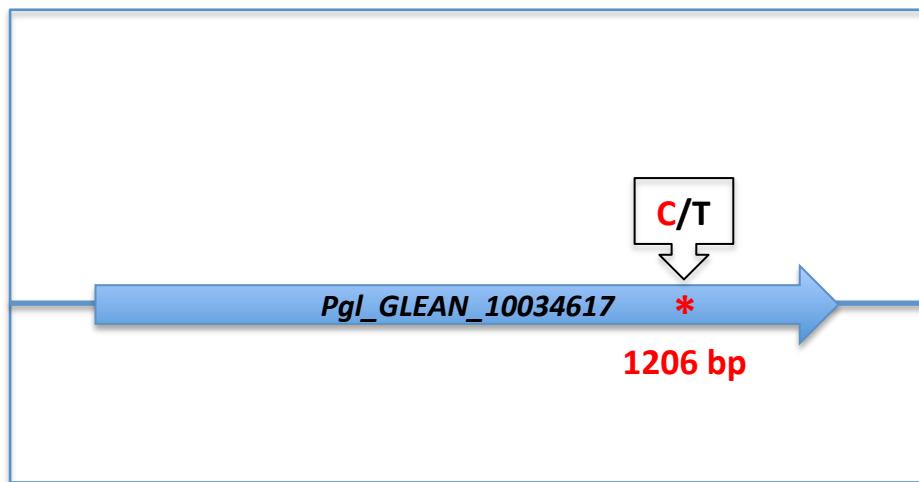
# Genetic bases root growth



Hangzhou, November 2018



# Genetic bases root growth



Hangzhou, November 2018



# Genetic bases root growth

249 (short root) = 337 (long root)

Plant-Specific Histone Deacetylases HDT1/2 Regulate  
*GIBBERELLIN 2-OXIDASE2* Expression to Control  
Arabidopsis Root Meristem Cell Number <sup>OPEN</sup>

Huchen Li,<sup>a</sup> Jesús Torres-García,<sup>a</sup> David Latrasse,<sup>b,c</sup> Moussa Benhamed,<sup>c,d</sup> Stefan Schilderink,<sup>a</sup> Wenkun Zhou,<sup>e</sup> Olga Kulikova,<sup>a</sup> Heribert Hirt,<sup>b,d</sup> and Ton Bisseling<sup>a,\*</sup>  
<sup>a</sup> Department of Plant Sciences, Laboratory of Molecular Biology, Wageningen University, 6706 PB Wageningen, The Netherlands  
<sup>b</sup> Unité de Recherche en Génomique Végétale, UMR INRA 1165, Université d'Evry Val d'Essonne, ERL CNRS 8196, Saclay Plant Sciences, 91057 Evry, France  
<sup>c</sup> Institut de Biologie des Plantes, CNRS-Université Paris-Sud 11, UMR 8618, 91405 Orsay cedex, France  
<sup>d</sup> King Abdullah University of Sciences and Technology, Thuwal 23955, Saudi Arabia  
<sup>e</sup> Department of Plant Sciences, Plant Developmental Biology, Wageningen University, 6706 PB Wageningen, The Netherlands

249 (short root) >> 337 (long root)

Progressive Inhibition by Water Deficit of Cell Wall Extensibility and Growth along the Elongation Zone of Maize Roots Is Related to Increased Lignin Metabolism and Progressive Stelar Accumulation of Wall Phenolics<sup>1</sup>

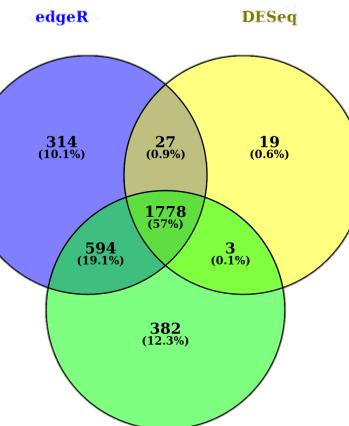
Ling Fan, Raphael Linker, Shimon Gepstein, Eiichi Tanimoto, Ryoichi Yamamoto, and Peter M. Neumann\*  
Plant Physiology Laboratory, Department of Environmental, Water, and Agricultural Engineering, Faculty of Civil and Environmental Engineering (L.F., R.L., P.M.N.) and Faculty of Biology (S.G.), Technion-Israel Institute of Technology, Haifa 32000, Israel; Plant Physiology Laboratory, Department of Information and Biological Sciences, Graduate School of Natural Sciences, Nagoya City University, Nagoya 467-8501, Japan (E.T.); and Biology and Chemistry Laboratory, Tezukayama University, Nara 631-8585, Japan (R.Y.)

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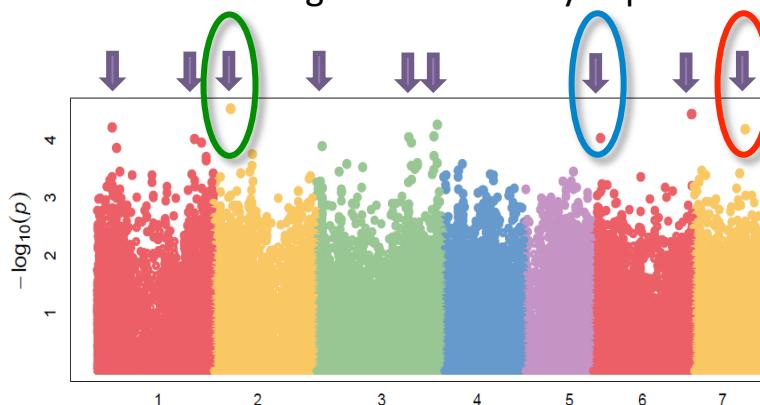
The Plant Cell, Vol. 23: 1107–1123, March 2011, www.plantcell.org © 2011 American Society of Plant Biologists

Patatin-Related Phospholipase pPLAIIβ-Induced Changes in Lipid Metabolism Alter Cellulose Content and Cell Elongation in *Arabidopsis* <sup>GW</sup>

Maoyin Li,<sup>a,b</sup> Sung Chul Bahn,<sup>a,b</sup> Liang Guo,<sup>a,b</sup> William Musgrave,<sup>b</sup> Howard Berg,<sup>b</sup> Ruth Welti,<sup>c</sup> and Xuemin Wang<sup>a,b,\*</sup>  
<sup>a</sup> Department of Biology, University of Missouri, St. Louis, Missouri 63121  
<sup>b</sup> Donald Danforth Plant Science Center, St. Louis, Missouri 63132  
<sup>c</sup> Kansas Lipidomics Research Center, Division of Biology, Kansas State University, Manhattan, Kansas 66506



RNAseq data from root tips of two contrasting lines revealed 1778 genes differently expressed



- ~ 100 expressed genes in QTL regions
- 3 SNPs located within genes with predicted functions for root growth



# Summary

- Pearl millet – crop for African agriculture adaptation to future climates
- Root growth - adaptative trait for abiotic stresses tolerance in pearl millet
- GWAS - QTLs for root growth (9) & rhizosheath formation (5)
- RNAseq on root tip of contrasted lines



# Perspectives



C. de la Fuente

## ➤ Validate/refine QTLs

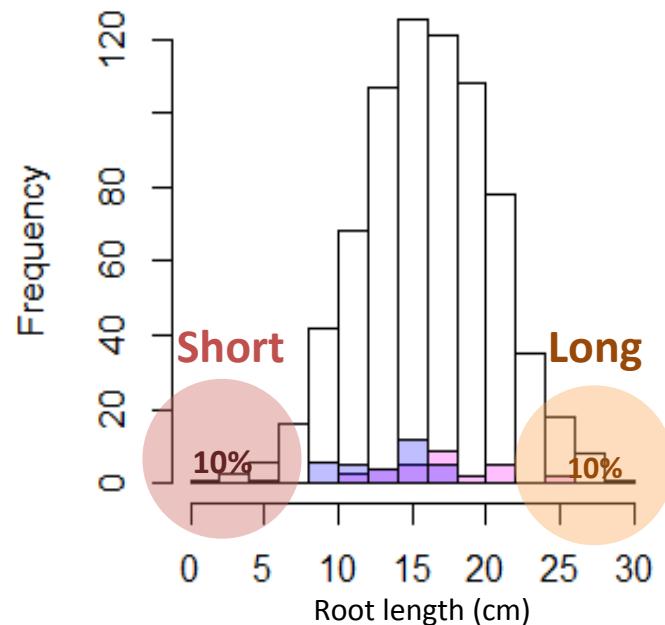
Slow growth parent

X

Fast growth parent

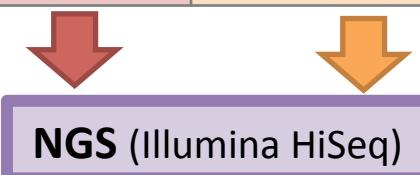


- 933 F2 seedlings 337x249 cross →
- 1116 F2 seedlings 109x57 cross →



Bulks Mean root length values ( $\pm$  SD) in cm

Cross	Short seminal	Long seminal
337 x 249	$8.15 \pm 1.84$ cm	$23.70 \pm 1.73$ cm
109 x 57	$6.86 \pm 1.75$ cm	$22.50 \pm 1.63$ cm





# Perspectives



C. de la Fuente

## ➤ Validate/refine QTLs

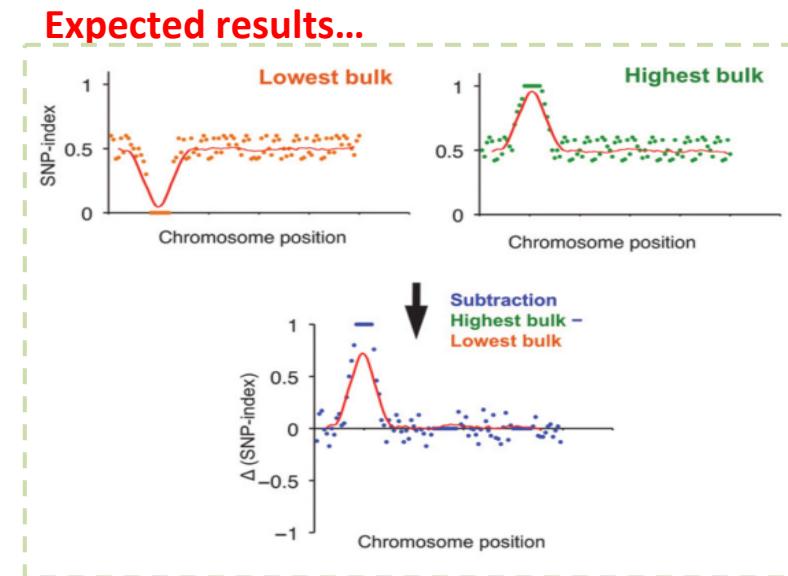
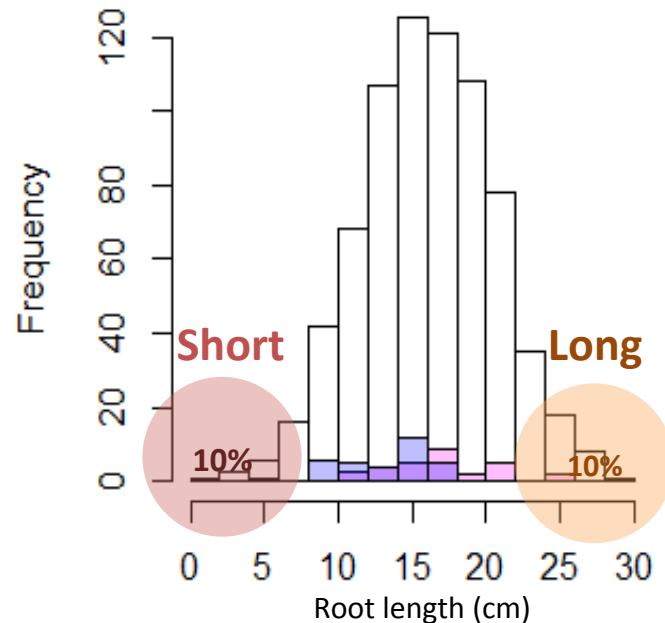
Slow growth parent

X

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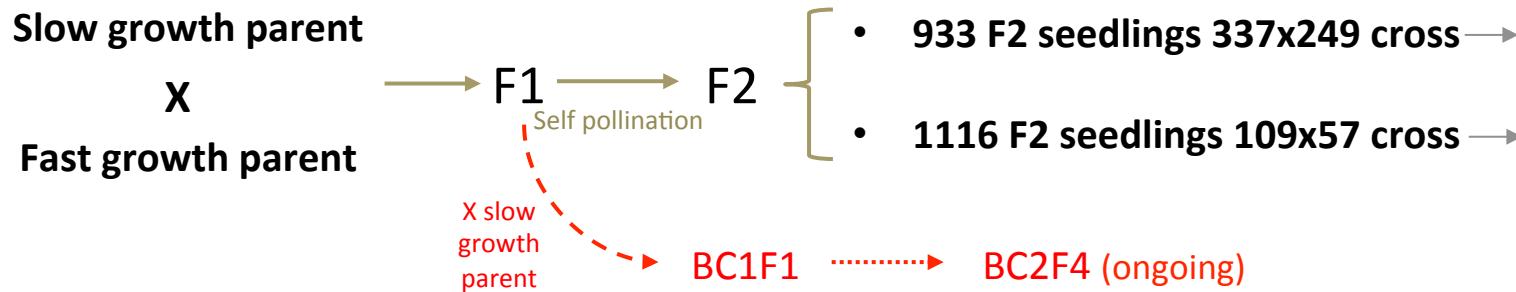
Adapted from: Takagi et al. (2013) The Plant Journal, 74: 174-183

Hangzhou, November 2018



# Perspectives

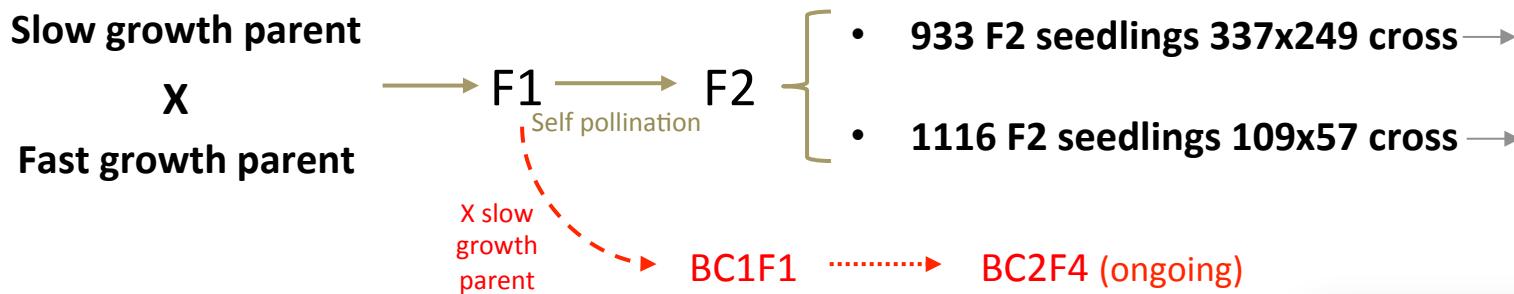
➤ Validate/refine QTLs





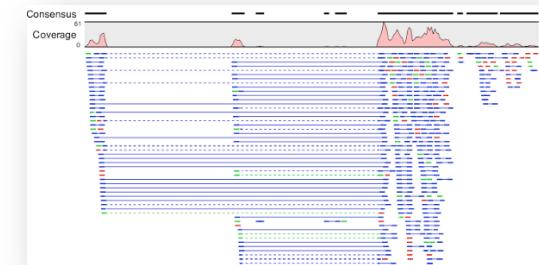
# Perspectives

- Validate/refine QTLs



- Refine gene annotation in QTL regions

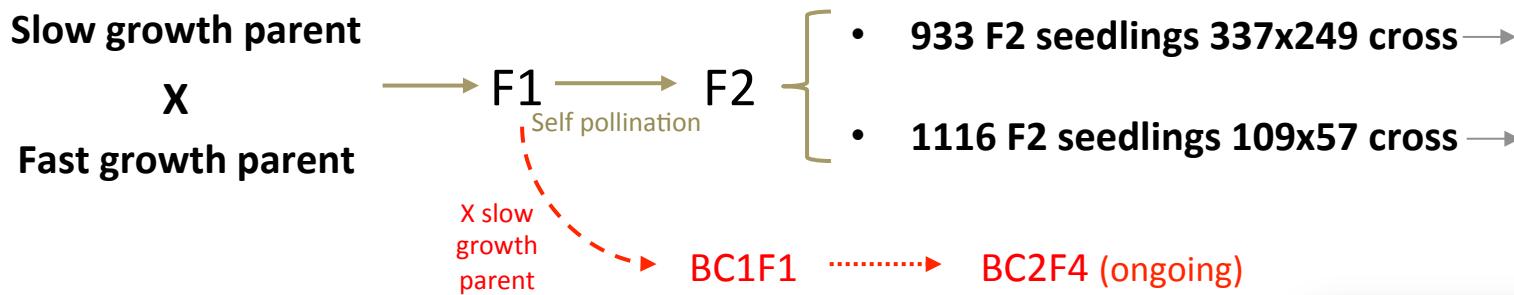
- RNAseq Data
- smallRNAseq





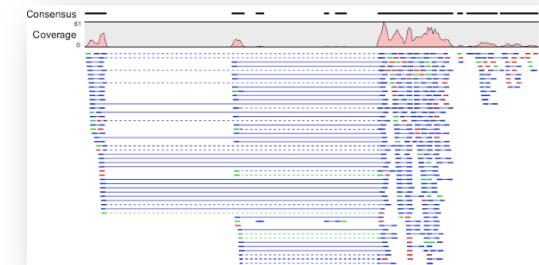
# Perspectives

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- Refine gene annotation in QTL regions

- RNAseq Data
- smallRNAseq

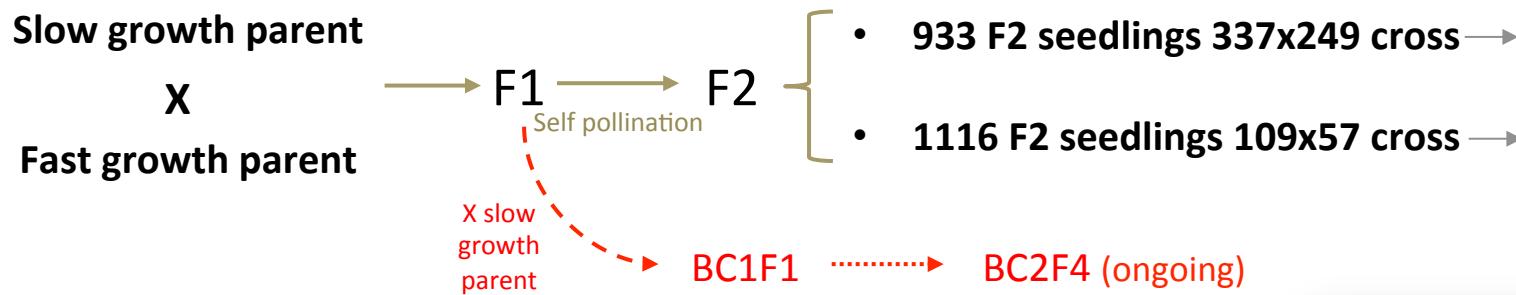


- Functional analysis of candidate genes



# Perspectives

- Validate/refine QTLs



- Refine gene annotation in QTL regions

- RNAseq Data
- smallRNAseq



- Functional analysis of candidate genes

- Measure impact validated QTL in field conditions

# Acknowledgments



**L. Laplaze**  
**Y. Vigouroux**  
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L. Zekraoui  
M. Couderc  
D. Moukouanga  
**M. Debieu**  
**C. de la Fuente-Canto**  
**S. Passot (PhD)**  
A. Ndour (PhD)  
A. Faye (PhD)  
M.-T. Mofini (PhD)  
F. Gnacko (MSc)



L. Cournac  
D. Masse  
C. Clermont  
L. Lardy  
K. Assigbetsé<sup>1</sup>  
M. Gueye  
M. Sitor (PhD)  
D. Tine



T. Heulin  
W. Achouak  
M. Barakat  
P. Ortet



N. Kane  
**B. Sine**  
A. Kane  
A. Diedhiou



P. Gangashetty  
C.T. Hash



M. Bennett  
D. Wells  
J. Atkinson  
C. Sturrock  
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