



OPEN

Linkage mapping and QTL analysis of flowering time in faba bean

David Aguilar-Benitez¹, Inés Casimiro-Soriguer¹, Fouad Maalouf² & Ana M. Torres¹  

Flowering time marks the transition from vegetative to reproductive growth and is key for optimal yield in any crop. The molecular mechanisms controlling this trait have been extensively studied in model plants such as *Arabidopsis thaliana* and rice. While knowledge on the molecular regulation of this trait is rapidly increasing in sequenced galeoid legume crops, understanding in faba bean remains limited. Here we exploited translational genomics from model legume crops to identify and fine map QTLs linked to flowering time in faba bean. Among the 31 candidate genes relevant for flowering control in *A. thaliana* and *Cicer arietinum* assayed, 25 could be mapped in a segregating faba bean RIL population. While most of the genes showed conserved synteny among related legume species, none of them co-localized with the 9 significant QTL regions identified. The *FT* gene, previously implicated in the control of flowering time in numerous members of the temperate legume clade, mapped close to the most relevant stable and conserved QTL in chromosome V. Interestingly, QTL analysis suggests an important role of epigenetic modifications in faba bean flowering control. The new QTLs and candidate genes assayed here provide a robust framework for further genetic studies and will contribute to the elucidation of the molecular mechanisms controlling this trait.

With its global production of 4.92 million metric tons¹ grown in 2.51 million ha, faba bean (*Vicia faba* L.) is the fourth most widely grown cool season legume after pea (*Pisum sativum*), chickpea (*Cicer arietinum*) and lentil (*Lens culinaris*). Due to its valuable content of protein and energy, faba bean is used as a livestock feed in both developing and developed countries, and as staple food in developing regions, especially in North and East Africa. Moreover, faba bean contributes to sustainable agriculture and plays an important role in the management of soil fertility through crop rotation and nitrogen fixation. Nevertheless, environmental conditions together with pests, diseases and weeds constrain the production of this major food crop.

Faba bean plants are particularly susceptible to frost, high temperature and/or low moisture during floral development and anthesis². Therefore, the time of flowering is a key trait in faba bean breeding in order to minimize exposure to critical stress and to produce novel varieties that are better adapted to local environments³. While two environmental factors, photoperiod and temperature, play a critical regulatory role in plant flowering, the response to these factors varies considerably among genotypes. Faba beans accessions are classified as, in general, day-neutral or long-day adapted, but photoperiod-unresponsive genotypes have also been identified^{4–8}.

Flowering time has been studied for decades both in model and crop plants⁹. A long list of regulatory genes have been identified in *A. thaliana* whose orthologues have also been identified in legume crops such as *Glycine max*¹⁰ or *Pisum sativum*¹¹. Despite some examples of gene loss or duplication, the main flowering genes are well catalogued and largely conserved among legumes^{10,12–14}.

Floral initiation depends on a system where photoreceptors perceive changes in daylength (photoperiod) and trigger the plant responses. Shim¹⁵ divided the photoperiodic flowering mechanisms into three steps: light input, circadian clock and output. Light information is integrated into innate photoperiodic timing mechanisms governed by the circadian clock to induce genes that trigger flowering. In legumes different classes of photoreceptors have been identified: three phytochromes (*PHYA*, *PHYB* and *PHYE*) and two cryptochromes (*CRY1* and *CRY2*)^{12,16}. In addition to the light signals, internal pathways convey information via the *FKF1/ZTL* gene family¹⁷. All these signals converge on the circadian clock, which is controlled by the transcription factors *CIRCADIAN CLOCK ASSOCIATED 1* (*CCA1*) and *LATE ELONGATED HYPOCOTYL* (*LHY*), *TIMING OF CAB 1* (*TOC1*), *EARLY FLOWERING 3* (*ELF3*) and *GIGANTEA* (*GI*). In long day (LD) plants, such as *Arabidopsis* or galeoid legumes, light signals promote the expression of *CONSTANS* (*CO*), which activates *FLOWERING LOCUS T* (*FT*) and *SUPPRESSOR OF OVEREXPRESSION OF CO1* (*SOC1*) to promote flowering at the meristem¹⁸ (Fig. 1). *FT* genes have an important position within the genetic hierarchy that controls flowering. *FT* belongs to the PEBP gene family, which includes five other member *TWIN SISTER OF FT* (*TSF*), *TERMINAL FLOWER 1* (*TFL1*), *A*.

¹Área de Genómica y Biotecnología, IFAPA Centro "Alameda del Obispo", Apdo 3092, 14080 Córdoba, Spain. ²International Center for Agricultural Research in the Dry Areas (ICARDA), Beirut, Lebanon. ✉email: anam.torres.romero@juntadeandalucia.es

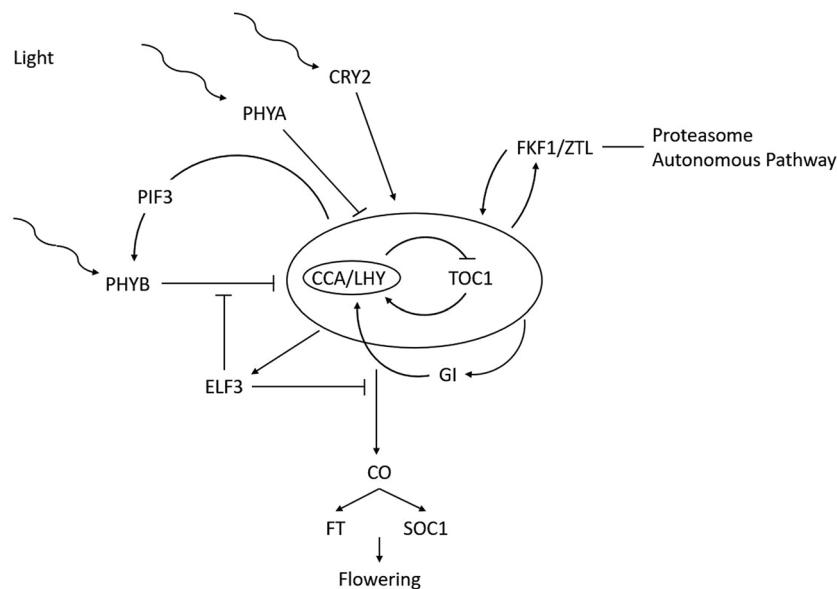


Figure 1. Schematic model of the genetic pathway of flowering time control by the photoperiod. The central circle represents the circadian clock. Modified from Mouradov et al. (2002).

thaliana CENTRORADIALIS homolog, MOTHER OF FT AND TFL1 (MFT), and BROTHER OF FT AND TFL1 (BFT)^{19–23}. TFL1 acts as a competitor of FT by competitive binding to FD²⁴, which in turn activates the floral meristem identity gene APETALA1 (API).

In legumes, the FT genes belong to three distinct subclades, FTa, FTb and FTc, each of which comprises several genes with distinct expression patterns in pea²⁵. The two key FT genes in leaves, FTa1 and FTb2, generate distinct mobile signals that influence flowering at the apex. FTb2 showed the highest induction under LD conditions in leaves, whereas FTa1 and FTc showed expression in apical buds. Hecht²⁵ proposed a model for the role and interactions of the pea FT genes, with FTa1 acting as a photoperiod-independent monitor of environmental variables that trigger the flowering, whereas FTb2 functions as the primary signal in leaves for promoting flowering in LD conditions.

Two legume crops stand out in flowering time studies, pea and soybean, with more than 20 loci identified in pea and at least 10 in soybean¹⁴. The majority of these loci may have been detected as QTLs in related legume crops but the correspondence is difficult to assess due to a lack of common markers and/or sequence information. For example, in chickpea or lentil, major QTLs corresponding to flowering time have been reported^{26–31}, but only in a few cases³², the orthology with other crops or model species has been assessed. In case of faba bean, just a few QTL analyses for flowering time have been reported^{33–36} but no information on the underlying genes and possible orthology, is currently available.

The aim of this study was to exploit translational genomics from model species or related legume crops to identify QTLs linked to flowering time in faba bean. For the first time, a set of genes relevant for flowering control in *A. thaliana*, *M. truncatula*, *P. sativum* and *C. arietinum* were assayed for polymorphisms and further mapped in a segregating faba bean RIL population. The objective was to saturate the map with candidate genes and to identify the QTLs that show a conserved location across different species. The results from this study will advance the understanding of the genetic basis of flowering time control and identify candidate genes and genomic regions of use for targeted molecular breeding of the trait in faba bean.

Results

Phenotypic evaluation. Three different flowering time traits were evaluated: DF1 (days from sowing to appearance of the first flower); DF50 (days until 50% of plants had a visible open flowers); and FL (flowering length, days between the start and the end of the flowering period). All three traits showed a continuous distribution, suggesting that they are controlled by multiple genes in this population (data not shown).

Mean phenotypic values and basic descriptive statistics of the traits are shown in Table 1. The two parental lines showed clear differences, DF1 and DF50 were shorter in Vf27 compared to Vf6, whereas FL was longer in 2007/08. The range of values for the three flowering traits in the RILs showed similar results across the years. DF1_2011/12 showed a higher mean value than in the previous years (DF1_2006/07 and DF1_2007/08) while the mean values in Lebanon for DF50 (DF50_2009/10 and DF50_2010/11) were lower than those recorded in Spain (DF50_2011/12).

The correlation analysis between the phenological traits DF1, DF50 and FL and years is summarized in Table 2. All correlations were significant (p value < 0.01) and the three traits displayed a high degree of intercorrelation. DF1 and FL in 2006/07 and 2007/08 showed high positive correlation (0.83 and 0.74, respectively), while strong negative correlations were observed between DF1 and FL in both seasons. This is due to trait compensation effects since the earlier the plants flower, the longer the flowering period lasts. A very high correlation value

Trait	Location	Vf6	Vf27	Range (min–max)	Mean ($\pm$ SE)
DF1_2006/07	Córdoba, Spain	108	98	87–131	105.8 $\pm$ 0.88
DF1_2007/08	Córdoba, Spain	102	82	79–125	100.6 $\pm$ 0.96
DF1_2011/12	Córdoba, Spain	–	–	101–137	120.6 $\pm$ 0.91
DF50_2009/10	Terbol, Lebanon	123	105	101–135	115.5 $\pm$ 1.01
DF50_2010/11	Terbol, Lebanon	131	115	105–133	120.6 $\pm$ 0.64
DF50_2011/12	Córdoba, Spain	–	–	104–144	130 $\pm$ 0.94
FL_2006/07	Córdoba, Spain	–	57	35–70	53.07 $\pm$ 0.84
FL_2007/08	Córdoba, Spain	48	57	20–66	43.79 $\pm$ 0.92

Table 1. Phenotypic values (mean $\pm$ SE) of days to first flower (DF1), days to 50% of flowering (DF50) and flowering length (FL) for parental and RILs from the cross Vf6 $\times$ Vf27 in each location and agronomic season.

	DF1_2006/07	DF1_2007/08	DF1_2011/12	DF50_2009/10	DF50_2010/11	DF50_2011/12	FL_2006/07	FL_2007/08
DF1_2006/07	1.00							
DF1_2007/08	0.83	1.00						
DF1_2011/12	0.55	0.50	1.00					
DF50_2009/10	0.67	0.67	0.64	1.00				
DF50_2010/11	0.45	0.49	0.51	0.86	1.00			
DF50_2011/12	0.63	0.55	0.94	0.66	0.52	1.00		
FL_2006/07	– 0.93	– 0.78	– 0.51	– 0.61	– 0.57	– 0.58	1.00	
FL_2007/08	– 0.76	– 0.96	– 0.49	– 0.60	– 0.43	– 0.52	0.74	1.00

Table 2. Flowering time traits correlation values. All the correlations showed a significant p value (< 0.01).

was also found between DF1_2011/12 and DF50_2011/12 (0.94) and between the two evaluations performed in Terbol, Lebanon (DF50_2009/10 and DF50_2010/11).

The ANOVA results for the genotype by environment ($G \times E$) analyses of each phenological trait are shown in Supplementary Table 1. The analysis revealed significant effects for genotypes (RILs), environment (years) and $G \times E$ interactions, identifying $G \times E$ interactions as an important factor in flowering time variation across genotypes and years.

Identification of flowering-related genes and primer design. We selected 31 flowering-related genes from *A. thaliana* (20 genes) and *chickpea* (11 genes) to study their implication in the control of flowering time in faba bean. Protein sequences were used in a BLASTp search to identify orthologous sequences in three legume species close to faba bean, *C. arietinum*, *M. truncatula* and *P. sativum*. Positive blast matches are summarized in Supplementary Table 2. Interestingly, the BLASTp search with *A. thaliana* CRY2 failed to return a positive match in *C. arietinum* and *M. truncatula*.

The sequences of these orthologues were used for designing primers (see materials and methods for details). Information on the primer sequences, size of the amplified genomic fragments and annealing temperatures for each candidate gene are shown in Supplementary Table 3. When applied to faba bean accessions, 28 out of 31 markers yielded amplification products. However, markers Vf_PM3, Vf_SPL1 and Vf_TICb from *C. arietinum* failed to amplify in faba bean.

The amplified sequences from the two parental lines were aligned to search for polymorphisms. Markers Vf_COP1 and Vf_FT produced DNA fragments of different length, which could be directly discriminated by agarose gel electrophoresis. These amplified length polymorphisms (ALPs) are derived from insertions or deletions yielding a different banding pattern. Markers Vf_PIF3 and Vf_GI did not show polymorphisms between the parental lines, while the remaining 24 markers (16 from *A. thaliana* and eight from *C. arietinum*) all revealed SNP polymorphisms. In 18 of these markers, the SNP was associated with a restriction site and thus these markers were transformed into CAPs. For the remaining six markers internal primers were designed. If the internal primer discrimination failed (Vf_TT8, Vf_FKF1 and Vf_PHYA), the SNPs were genotyped using the MassArray iPLEX (Sequenom) platform (<http://www.cegen.org>). Only Vf_ELF4 could not be genotyped using any of the genotyping techniques described.

Genetic mapping. The genotypic data obtained with the flowering time candidate markers were combined with the data set previously reported in this RIL population^{33,37}. Information on the map positions is shown in Table 3. All the new markers fit the expected 1:1 Mendelian ratio except for Vf_FT which showed a distorted segregation skewed towards the Vf6 allele (Table 3). The linkage map, spanning 4.184 $\square$ cM, consists of six main linkage groups and five smaller arrays (Fig. 2). The existence of common markers with previous consensus

Marker	X ² (1:1)	P	Vf_Chr	Ca_Chr	Mt_Chr	Ps_LG
Vf_EIN4	0.59	0.442	3	4	1	Scaffold
Vf_GA20ox	2.086	0.149	3	4	1	2
Vf_GA20ox3	2.922	0.087	3	4	1	2
Vf_ICCM	0.011	0.916	3	4	1	2
Vf_TICa	3.658	0.056	3	4	1	2
Vf_TT8	0.385	0.535	3	4	1	2
Vf_LHY	0	1	2	3	7	5
Vf_CO	1.316	0.251	1	4	7	5
Vf_COP1	0.91	0.34	4	2	5	1
Vf_CRY1	0.976	0.323	1	2	5	1
Vf_CRY2	0.05	0.823	3	–	–	2
Vf_ELF3	0.047	0.829	2	5	3	3
Vf_FD	0.419	0.518	1	8	5	1
Vf_FKF1	0.235	0.628	6	6	8	6
Vf_FT	26.385	0*	5	3	6	6
Vf_TFL1	0.400	0.527	5	3	7	5
Vf_LFY	1.754	0.185	2	5	3	3
Vf_LUX	1.136	0.286	6	6	4	7
Vf_PHYA	1.704	0.192	3	4	1	2
Vf_PHYB	0.117	0.732	1	1	2	6
Vf_PHYE	0.59	0.442	1	1	2	6
Vf_SOC1	0.048	0.827	4	3	8	–
Vf_SVP	1.988	0.159	1	Scaffold	5	1
Vf_TEM	0.049	0.825	1	Scaffold	5	1
Vf_TOC1	0.762	0.383	4	7	4	4

Table 3. Genetic segregation, chi-square test and location of the flowering time gene markers in faba bean (Vf) and related legume species: Ca, *Cicer arietinum*; Mt, *Medicago truncatula* and Ps, *Pisum sativum*). Bold values highlight syntenic chromosomes (Chr)/and linkage groups (LGs). Lack of fit to the expected 1:1 segregation is indicated by an asterisk.

maps^{34,38} allowed us to assign all the LGs to the corresponding faba bean chromosomes (chr.). All candidate-gene markers were included in the new genetic map, with numbers ranging from one in LGVI to six in LGI and LGIII.

We next checked whether the marker genes share syntenic positions in *V. faba* and the closely related species *M. truncatula*, *P. sativum* and *C. arietinum* (Table 3). The level of synteny was quite high (69%), with 18 out of 25 gene markers conserved in at least one other species. Twelve *V. faba* genes were syntenic with *C. arietinum*, 15 with *M. truncatula* and 16 with *P. sativum*. Finally, 12 markers (Vf_GA20ox, Vf_GA20ox3, Vf_ICCM0293, Vf_TICa, Vf_TT8, Vf_CRY1, Vf_ELF3, Vf_FD, Vf_TFL1, Vf_LFY, Vf_LUX and Vf_PHYA) showed conserved synteny in all four species tested, preserving co-localization of these genes in their homologous chromosomes.

QTL analysis. We identified 12 significant QTLs (LOD threshold > 3.3) related with flowering time in chr. I, II, III, IV and V (Table 4, Fig. 2). Except for DF1_07-08(1), DF1_07-08(2), DF50_11-12(1) and FL_06-07(3) the additive effects of the associated markers were positive, indicating that most early flowering alleles originate from the Vf27 parental line.

We identified 4 significant QTLs for DF1. Two of these, DF1_06-07(2) and DF1_07-08(3), with a LOD peak > 5, explaining 11.1 and 17.4% of the phenotypic variance, respectively, colocalized in chr. V associated with the *Medicago truncatula* (Mtr) marker Mtr7g084800. In addition, DF1_07-08(3) showed a second significant peak that colocalized with FL_07-08(3) and explained 11.1% of the variation. The other two QTLs were located in chr. II (DF1_07-08(1)) and IV (DF1_07-08(2)) and explained 10.7 and 11% of the trait variation.

For DF50 scored in 2011/12, three significant QTLs were found in chr. I, II and V. The largest main effect QTL was DF50_11-12(2), which explained 17.9% of the phenotypic variance and is located in chr. II flanked by GAI18 and Vf_Mtr3g096560.

Finally, 5 significant QTLs for FL were detected in chr. I, III, IV and V. Two of these colocalized with other DF1 QTLs in chr. IV (FL_07-08(2)) and chr. V (FL_06-07(3)) (Fig. 2). Moreover, two additional QTLs for FL_2006/07 were detected in chr. I (with a second peak between Mtr4g021350 and Mtr5g097910) and chr. III (with a second peak next to Mtr1g087900(LG38)), explaining 7.4 and 11.6% of phenotypic variation, respectively.

Four medium-sized QTLs failed to reach the defined threshold: FL_07-08(1) in chr. II, DF1_06-07(1) in chr. IV and DF1_11-12(1) and DF50_09-10(2) in chr. V. Nevertheless, these QTLs are displayed in Fig. 2 and Table 4 since they co-localized with other significant QTLs. However, none of the 25 flowering genes included in the

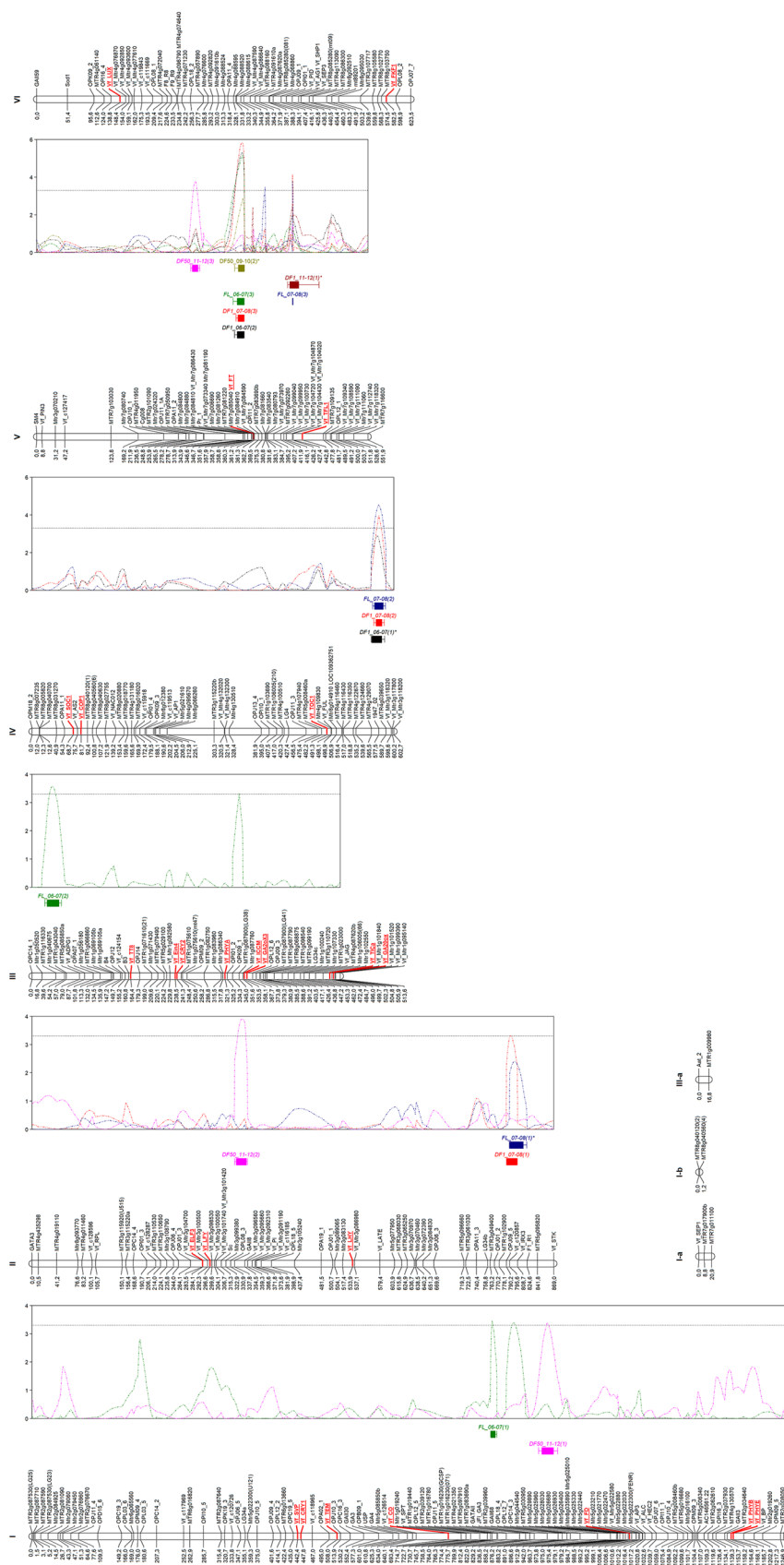


Figure 2. Linkage map and QTLs for flowering time traits detected in the Vf6 × Vf27 RIL population. QTL locations are represented with bars (2-LOD interval) and boxes (1-LOD interval). Candidate gene markers included in the map are highlighted in red. DF1: days to first flower, DF50: days to 50% of flowering, FL: flowering length. No significant QTLs were identified with *.

Trait	Peak	Chr	Flanking markers	LOD	Additive effects	R ²
FL_06-07(1)	766.313	I	OPJ11_5	3.46	2.27494	7.4
DF50_11-12(1)	858.208	I	Mtr2g039960	3.39	− 2.91921	9.1
DF50_11-12(2)	349.829	II	GAI18/Vf_Mtr3g096560	3.91	3.90166	17.9
DF1_07-08(1)	796.562	II	Vf_c120857	3.31	− 3.06379	10.7
FL_07-08(1)*	805.562	II	Vf_IRX3	2.39	2.4726	7.6
FL_06-07(2)	33.823	III	Mtr1g116330	3.57	2.86524	11.6
DF1_06-07(1)*	577.528	IV	1947_02	2.75	− 2.10807	5.3
DF1_07-08(2)	577.528	IV	1947_02	3.98	− 3.16022	11
FL_07-08(2)	577.528	IV	1947_02	4.54	3.17776	12.6
DF50_11-12(3)	265.522	V	Mtr7g024320	3.72	3.36068	13.2
DF1_06-07(2)	343.911	V	Mtr7g084800	5.26	3.85206	11.1
DF1_07-08(3)	343.911	V	Mtr7g084800	5.76	4.90158	17.4
DF50_09-10(2)*	343.911	V	Mtr7g084800	2.86	4.53734	12.8
FL_06-07(3)	343.911	V	Mtr7g084800	5.19	− 3.41023	12.2
DF1_11-12(1)*	426.159	V	Vf_Mtr7g104720/Vf_Mtr7g104870	2.57	− 3.22325	10.6
FL_07-08(3)	426.167	V	Vf_Mtr7g104870	3.80	3.29528	11.1

Table 4. QTLs for flowering time traits identified in the Vf6 × Vf27 RIL population. *No significant QTL.

map appear to be plausible candidates for the observed QTLs, and only FT and TFL1 (in chr. V) were located close or within the confidence interval of a significant QTL.

In summary, chromosome V accumulates most of the consistent and stable flowering time QTLs detected across different locations and years. These QTLs were distributed in two regions. The first including QTLs for DF1, DF50 and FL is located close to the marker Mtr7g084800, while the second including QTLs for DF1 and FL is close to Vf_Mtr7g104870. Additional relevant genomic regions were identified in chr. II and IV, where DF1 and FL QTLs colocalized in the distal part of both chromosomes and explained between 10.7–17.9% and 11–12.6% of the phenotypic variation, respectively.

To identify the orthologous genes flanking the QTLs, we performed a BLASTp search against the *Arabidopsis* protein database in the NCBI platform. These results are shown in Supplementary Table 4.

Discussion

Flowering time is one of the most important adaptive traits in plants³⁹ and its regulation crucially affects crop yield. As many other temperate legume crops, faba bean synchronizes flowering to match the seasons by monitoring cues such as temperature and photoperiod. Thus, exposure to winter cold (vernalization) results in the competence to flower during the following spring in response to increasing day length signals.

Both the vernalization pathway and the photoperiod pathway have been characterized in detail for the model plant *A. thaliana*, where more than 100 genes contributing to the flowering time control have been identified⁴⁰. Most of the key genes and gene families of the *Arabidopsis* flowering time pathway appear to be conserved in legumes, with a few of them having undergone duplications (e.g. *CRY2*) or loss (e.g. *PHYC*)¹⁴. Thus, there is significant potential for translating the knowledge on flowering control from the model or related legume species to faba bean.

Here we conducted a comprehensive analysis of the genetic determination of flowering time in faba bean by systematically evaluating three related traits in a segregating RIL population across different locations and years using a combination of comparative genomics, candidate gene mapping and QTL analysis. We selected a set of 31 genes controlling flowering time in *Arabidopsis* and chickpea to fine map faba bean target regions and identify genes co-localizing with previously identified QTLs, and were able to genotype and include 25 of them in the faba bean linkage map. Most of these markers showed a conserved synteny with the related legume crops *Medicago*, *Pisum* and *Cicer* (Table 3), although none of them co-localized with previously reported faba bean QTLs. Importantly, only *FT* gene mapped close to the most prominent and consistent QTL region (see below).

The analysis identified 12 significant QTLs for flowering time. The most important region in chr. V, contained five QTLs for different flowering traits that were stable and consistently detected across different years. The co-localization of two putative QTLs, DF1_11-12(1) and DF50_09-10(2), which did not reach the established significance level, further corroborates the importance of this genomic region flowering time control.

This region on chr. V was first identified by Cruz-Izquierdo³³, with a major QTL near markers Pis_GEN_6_3_1 (Mtr7g084800) and AnMtS37 (Mtr7g081220). Subsequent studies in faba bean^{35,36}, showed a relation of days to flowering with the SNP marker Vf_Mt7g084010, which mapped close to Mtr7g084800. Our present results confirm that Mtr7g084800 is the marker mapping closest to the most relevant QTL peak (Table 4, Fig. 2). Mtr7g084800 corresponds to a photosynthetic glyceraldehyde-3-phosphate dehydrogenase (GAPDH), which participates in the Calvin cycle, is transcriptionally regulated by light⁴¹ and highly expressed in leaves and floral stalks⁴².

Studies in temperate legume species such as *M. truncatula*, chickpea, narrow-leaved lupin, alfalfa and *Lotus japonicus* confirmed the conservation of this major flowering time QTL in a region syntenic with a section of

Medicago chr. 7, containing a tandem array of *FTa* and *FTC* genes (reviewed by Weller and Ortega¹⁴). As mentioned above, we found that *FT* was the only candidate gene mapping close to this QTL region, where marker Mtr7g084800 is surrounded by a cluster of *FT* flowering time loci (Supplementary Table 5). We attempted to design primers pairs corresponding to the different *FT* candidate genes present in the *M. truncatula* QTL interval⁴³ but the high sequence conservation prevented further amplification. Using the available *M. truncatula*, *P. sativum* and *V. faba* *FT* sequences, we were able to design primer pairs for Mtr7g085040 (*FTC*) as well as for a second *V. faba* *FT*, although linkage analysis mapped both markers to the same position confirming the high sequence conservation. Because none of the new *FT* markers genotyped was directly linked to the QTL found in chr. V, the identity of the responsible gene(s) remains to be determined.

Also, in chr. V, marker Vf_Mtr7g104870 was associated with QTL FL_07-08(3) and DF1_11-12(1), although the latter did not reach the significance level. Vf_Mtr7g104870 shows homology with the *A. thaliana* AT4G18590 gene encoding one of the three subunits of DNA Replication Protein A (RPA). RPA is a protein complex with a critical function in DNA replication, repair, recombination and epigenetic maintenance. These proteins play important roles in epigenetic gene silencing and in the regulation of the meristem development in *A. thaliana*⁴⁴. Interestingly, vernalization in *Arabidopsis* involves downregulation of *FLC*, primary floral repressor^{45,46}. Bastow⁴⁷ showed that vernalization causes histone methylation in the *FLC* gene, thereby preventing the transition to the reproductive state.

Further, we detected QTLs in chr. I, II, III and IV. The QTL DF50_11-12(1) in chr. I was associated with marker Mtr2g039960, which corresponds to eukaryotic translation initiation factor 4A1 (*EIF4A1*) according to a BLASTp search against the *A. thaliana* genome database (Supplementary Table 3). *EIF4A1* was previously shown to modulate the light/dark cycle in mature leaves⁴⁸. Interestingly, *A. thaliana* mutants in *EIF4A1* showed a pronounced late-flowering phenotype and significantly altered expression of *FLC* and *SOC1*, a positive and negative regulator of *FT* genes, respectively⁴⁹. Mutation in another *EIF4* gene (*EIF4G*) of *A. thaliana* resulted in a late flowering phenotype⁵⁰. In maize, *EIF4A1* was also described as a candidate gene for flowering time⁵¹.

In chr. II we detected QTL DF1_07-08(1), which colocalizes with FL_07-08(1), although it did not reach the significance level. Both are associated with markers Vf_c120857 and Vf_IRX3. Vf_c120857 (Mtr3g449590) shows homology with AT4G23820, a pectin lyase-like superfamily protein, which is apparently unlinked to flowering. A second QTL in chr. II, DF50_11-12(2), is flanked by markers GAI18 and Vf_Mtr3g096560. While GAI18 is a microsatellite, Vf_Mtr3g096560 corresponds to *A. thaliana*, DNA polymerase delta (δ) subunit 1 (AT5G63960), which is part of a complex of three DNA polymerases. DNA polymerase α (Pol α) initiates DNA strand synthesis while Pol ϵ and Pol δ perform the synthesis in the leading and lagging strands, respectively. In *A. thaliana*, Pol δ was reported to regulate flowering via epigenetic marks⁵². According to the proposed model Pol δ is delayed at high temperatures, triggering a DNA replication stress response which results in methylation of a H3K4me3 histone in the *SEPALLATA3* (*SEP3*) locus. This epigenetic mark promotes transcription *SEP3*, which participates in a feedback loop with *FT* genes, thus linking Pol δ activity to the establishment of transcriptional epigenetic marks affecting flowering. These findings are in line with the fact that in *Arabidopsis* *esd7* and *abo4-1* mutants, discrete regions at the *FLOWERING LOCUS T* (*FT*) are enriched in H3K4me3, correlating with higher *FT* and lower *FLC* expression levels⁵³.

The QTL FL_06-07(2) in chr. III was associated with Mtr1g116330, a PRLI-interacting factor specifically expressed during flower development in *Acacia mangium*⁵⁴. In rice, genetic analysis and fine mapping of a set of complementary genes controlling late heading (flowering time) also identified a PRLI-interacting factor as a putative candidate for LH1⁵⁵.

Finally, consistent QTLs for DF1 and FL co-localized in chr. IV were associated with the 1947_02 marker corresponding to the *M. truncatula* Mtr4g132540 gene, a homolog of the jumonji domain-containing protein JmjC in *A. thaliana* (Supplementary Table 3). JmjC proteins regulate epigenetic processes involved in plant growth, development and disease resistance^{56–58}. In *A. thaliana* several JmjC members have been characterized. They play a critical role in flowering time control via chromatin remodeling involving modifications of histones that either promote or repress gene expression. Members of the two known families of histone demethylases in eukaryotes, lysine-specific demethylase 1 (LSD1) and Jumonji C-terminal domains are involved in regulating both defense and developmental (flowering time) processes^{59,60}, as now is also suggested in faba bean.

The QTL analysis identified *EIF4A1* in chr. I and PRLI-interacting factor in chr. III, as well as three candidate genes involved in flowering time regulation through histone modifications, RPA in chr. V, DNA polymerase delta (δ) in chr. II and JmjC in chr. IV. This result highlights the important role of epigenetic modifications in faba bean flowering control. Based on this finding, we propose that conserved epigenetic pathways in plants can be exploited as a novel source to identify markers for flowering control and stress tolerance⁶¹.

The QTLs and candidate genes identified here provide a robust framework for further genetic studies aimed at elucidating the mechanisms controlling flowering time. The ongoing faba bean genome sequence assembly together with the development of dense, gene-based genetic maps will further facilitate fine mapping of relevant QTL regions. These QTLs need to be transferred to effective markers for further use in the breeding programs. Therefore, additional flowering time studies involving new faba bean genetic backgrounds and expression analyses will validate the usefulness of the identified candidates in flowering control. The predicted outcomes will provide targets for an effective selection of flowering traits in faba bean breeding programs.

Material and methods

Plant material. A recombinant inbred line (RIL) population of 124 individuals derived from the faba bean cross Vf6 $\times$ Vf27 was used in this work. Vf6 is an *equina* medium-seeded field bean with beige seed coat and late flowering time while Vf27 is a black and small-seeded *paucijuga* primitive form with earlier date of flowering,

with a phenotype similar to the hypothetical wild progenitor⁶². This RIL population was developed at IFAPA and well-studied in previous mapping and QTL research^{63–65}.

Phenotypic evaluation. Evaluations were conducted at IFAPA-station in Alameda del Obispo, Córdoba in Spain (37.51° N, 4.48° W, 94 m) during 2006/2007, 2007/2008 and 2011/2012 seasons) and at ICARDA, Terbol Research Station, Bekaa Valley in Lebanon (35.98° N, 33.88° E, 890 m) during 2009/2010 and 2010/2011 winter cropping seasons. The mean annual temperature in Córdoba is 24.6 °C and the annual rainfall is 536 mm, mostly concentrated from December to February. In Terbol, the mean annual temperature is 15.4 °C and the annual rainfall of 559 mm. A randomized complete block design with two replicates per genotype was used in both environments. The experimental unit consisted of 10 plants including RILs and parental lines. Sowing dates were during the first half of November in the five growing seasons.

Flowering traits were recorded using different methods: DF1, scored as the number of days from the sowing until the appearance of the first flower; DF50, recorded as the number of days until the 50% of the plants had visible open flowers and FL (flowering length), scored as the difference of days between the start and the end of flowering period (when no flowers were present in the plants). DF1 was evaluated in 2006/07, 2007/08 and 2011/12 (only in Spain); DF50 was evaluated in 2009/10, 2010/11 and 2011/12 (in Spain and Lebanon) and FL was evaluated in 2006/07 and 2007/08 (only in Spain). Evaluations from 2006/07 to 2007/08 were previously reported by Cruz-Izquierdo³³.

Global means, ranges, standard error (SE), histograms of frequency and statistical analyses were carried using the R software version 4.02⁶⁶.

Flowering-related genes and primer design. To develop molecular markers associated with flowering time in faba beans, we used a candidate gene-based strategy⁶⁷. Thirty-one flowering-related genes described in other species (mostly in chickpea and *A. thaliana*) were selected (Supplementary Table 2). Then using the BLAST tool, homologous sequences were identified in *Medicago truncatula* and *Pisum sativum*. Positive results were used as well to search for homologous sequences in faba bean transcriptome databases⁶⁸. The obtained sequences were aligned using the Geneious software suite (v. 7.1.9; Biomatters, Auckland, New Zealand) to identify conserved domains and design the primer pairs. When possible, primers were designed on faba bean sequences, otherwise, we used the sequence of related species.

A set of exonic primers flanking the intronic sequences with a size of 20–25 pb, GC contents of 45–60% and Tm of 60 ± 1 °C were designed to amplify fragments in a 300–1200 bp range. The markers developed in this work were named as the corresponding candidate gene preceded by Vf (Supplementary Table 3).

PCR amplification and polymorphism detection. Genomic DNA was isolated from young leaves using a CTAB protocol⁶⁹. PCR amplification consisted in 25 µl of reaction containing 4 ng template DNA, 1 × PCR buffer, 2 mM of MgCl₂, 0.4 mM of dNTPs, 0.2 µM of each primer and 1 unit of Taq polymerase (Biotools B&M Labs, Madrid, Spain). The amplification profile was an initial denaturation at 94 °C for 5 min, followed by 30 cycles of 45 s at 94 °C, 45 s at 56–60 °C and 45 s at 72 °C, with a final extension step of 8 min at 72 °C. PCR products were separated using 2% agarose gels. Positive amplifications were purified using a standard 3 M sodium acetate (pH 4.5) and ethanol protocol. The purified DNA was used for direct sequencing (STABVIDA, Portugal). Finally, the sequences from parental and contrasted RILs for flowering time were aligned to detect SNPs (Single Nucleotide Polymorphisms). Those SNPs with recognition sites for restriction enzyme digestion between lines were converted into CAPS makers (Cleaved Amplified Polymorphism Sequence) to genotype the whole RIL population. Restriction digestions were performed following the supplier's instructions and visualized in 2% agarose gels. For SNPs lacking restriction enzymes in the polymorphic site, internal primers were designed using the technique Tetra-Primer ARMS-PCR described by Medrano and Oliveira⁷⁰. When the internal primers did not yield amplification products or a clearly scorable segregation, the genotyping was carried out at CEGEN-PRB3-ISCHII (<http://www.cegen.org>) using the MassArray iPLEX (Sequenom) SNP genotyping platform from the Spanish National Genotyping Center facility of the University of Santiago de Compostela.

Genetic mapping and QTL analysis. The preceding maps developed in the Vf6 × Vf27 RIL population include approximately 425 RAPDs, ESTs and SNP markers^{33,37}. The new polymorphic flowering time markers were genotyped in this RIL population and incorporated into the previous dataset^{33,37} to develop a new linkage map. Chi-squared tests to determine goodness of fit to the expected segregation ratio of 1:1 were carried out for all markers segregating in the RIL population. Linkage analysis was performed using JoinMap v4.0⁷¹ with maximum likelihood option. Markers were grouped at a minimum LOD score of 3 and a maximum recombination fraction of 0.25 as general linkage criteria to establish linkage groups. Kosambi's function was applied to convert the recombination frequencies into genetic distances⁷².

QTL analysis was conducted by MapQTL v5.0 software⁷³. First, the nonparametric Kruskal–Wallis test was performed to estimate the marker significance level and the marker-phenotype association. Then, the interval mapping^{74,75} was applied to identify putative QTLs in each linkage group (LG). Markers significant at $p = 0.01$ were used as cofactors in the multiple QTL analysis (MQM)^{76–78}. QTL significance (p value) was determined by permutation analysis using 1000 replicates⁷⁹, as implemented in MapQTL 5.0. Only QTLs with a LOD higher than the p -value were declared as significant. MapChart software⁸⁰ was used to represent the QTLs confidence interval. The support intervals were defined as LOD-1 and LOD-2 around the maximum LOD of QTL. If QTLs for the same trait detected in different environments had overlapping 2-LOD support intervals, they were considered to be the same QTL and also been designated as consistent QTLs.

Ethical approval. Faba bean is a common crop extensively cultivated in the world. The authors declare that the experimental research work conducted in this study was carried out in accordance with relevant guidelines and regulations.

Data availability

The datasets generated and analyzed during the current study are available from the corresponding author on request.

Received: 4 December 2020; Accepted: 3 June 2021

Published online: 02 July 2021

References

1. FAOSTAT. Food and Agriculture Organisation of the United Nations (2018). <http://www.fao.org/faostat/en/#data/QC>. Accessed 23 Oct 2020.
2. Smith, M. L. Factors affecting flower abscission in field beans (*Vicia Faba* L. *Minor*). (Durham Theses, Durham University, 1982).
3. Jung, C. & Müller, A. E. Flowering time control and applications in plant breeding. *Trends Plant Sci.* **14**, 563–573 (2009).
4. Evans, L. T. Environmental control of flowering in *Vicia faba* L. *Ann. Bot.* **23**, 521–546 (1959).
5. Ellis, R. H., Summerfield, R. J. & Roberts, E. H. Effects of temperature, photoperiod and seed vernalization on flowering in faba bean *Vicia faba*. *Ann. Bot.* **61**, 17–27 (1988).
6. Ellis, R. H., Roberts, E. H. & Summerfield, R. J. Variation in the optimum temperature for rates of seedling emergence and progress towards flowering amongst six genotypes of faba bean (*Vicia faba*). *Ann. Bot.* **62**, 119–126 (1988).
7. Ellis, R. H., Summerfield, R. J. & Roberts, E. H. Flowering in faba bean: Genotypic differences in photoperiod sensitivity, similarities in temperature sensitivity, and implications for screening germplasm. *Ann. Bot.* **65**, 129–138 (1990).
8. McDonald, G. K., Adisarwanto, T. & Knight, R. Effect of time of sowing on flowering in faba bean (*Vicia faba*). *Aust. J. Exp. Agric.* **34**, 395 (1994).
9. Song, Y. H., Ito, S. & Imaizumi, T. Flowering time regulation: Photoperiod- and temperature-sensing in leaves. *Trends Plant Sci.* **18**, 575–583 (2013).
10. Kim, M. Y., Shin, J. H., Kang, Y. J., Shim, S. R. & Lee, S.-H. Divergence of flowering genes in soybean. *J. Biosci.* **37**, 857–870 (2012).
11. Weller, J. L. *et al.* Update on the genetic control of flowering in garden pea. *J. Exp. Bot.* **60**, 2493–2499 (2009).
12. Hecht, V. *et al.* Conservation of Arabidopsis flowering genes in model legumes. *Plant Physiol.* **137**, 1420–1434 (2005).
13. Watanabe, S., Harada, K. & Abe, J. Genetic and molecular bases of photoperiod responses of flowering in soybean. *Breed. Sci.* **61**, 531–543 (2012).
14. Weller, J. L. & Ortega, R. Genetic control of flowering time in legumes. *Front. Plant Sci.* **6**, 207 (2015).
15. Shim, J. S., Kubota, A. & Imaizumi, T. Circadian clock and photoperiodic flowering in Arabidopsis: *CONSTANS* is a hub for signal integration. *Plant Physiol.* **173**, 5–15 (2017).
16. Platten, J. D. *et al.* The cryptochrome gene family in pea includes two differentially expressed *CRY2* genes. *Plant Mol. Biol.* **59**, 683–696 (2005).
17. Imaizumi, T., Tran, H. G., Swartz, T. E., Briggs, W. R. & Kay, S. A. *FKF1* is essential for photoperiodic-specific light signalling in Arabidopsis. *Nature* **426**, 302–306 (2003).
18. Mouradov, A., Cremer, F. & Coupland, G. Control of flowering time: Interacting pathways as a basis for diversity. *Plant Cell* **14**(Suppl), S111–S130 (2002).
19. Bradley, D., Ratcliffe, O., Vincent, C., Carpenter, R. & Coen, E. Inflorescence commitment and architecture in Arabidopsis. *Science* **275**, 80–83 (1997).
20. Mimida, N. *et al.* Functional divergence of the *TFL1*-like gene family in Arabidopsis revealed by characterization of a novel homologue. *Genes Cells* **6**, 327–336 (2001).
21. Yoo, S. Y., Kardailsky, I., Lee, J. S., Weigel, D. & Ahn, J. H. Acceleration of flowering by overexpression of *MFT* (*MOTHER OF FT AND TFL1*). *Mol. Cells* **17**, 95–101 (2004).
22. Yamaguchi, A., Kobayashi, Y., Goto, K., Abe, M. & Araki, T. *TWIN SISTER OF FT* (*TSF*) acts as a floral pathway integrator redundantly with *FT*. *Plant Cell Physiol.* **46**, 1175–1189 (2005).
23. Yoo, S. J. *et al.* *BROTHER OF FT AND TFL1* (*BFT*) has *TFL1*-like activity and functions redundantly with *TFL1* in inflorescence meristem development in Arabidopsis. *Plant J.* **63**, 241–253 (2010).
24. Ahn, J. H. *et al.* A divergent external loop confers antagonistic activity on floral regulators *FT* and *TFL1*. *EMBO J.* **25**, 605–614 (2006).
25. Hecht, V. *et al.* The pea *GIGAS* gene is a *FLOWERING LOCUS T* homolog necessary for graft-transmissible specification of flowering but not for responsiveness to photoperiod. *Plant Cell* **23**, 147–161 (2011).
26. Fratini, R., Durán, Y., García, P. & de la Vega, M. P. Identification of quantitative trait loci (QTL) for plant structure, growth habit and yield in lentil. *Span. J. Agric. Res.* **5**, 348 (2007).
27. Aryamanesh, N., Nelson, M. N., Yan, G., Clarke, H. J. & Siddique, K. H. M. Mapping a major gene for growth habit and QTLs for ascochyta blight resistance and flowering time in a population between chickpea and *Cicer reticulatum*. *Euphytica* **173**, 307–319 (2010).
28. Kahriman, A., Yilmaz Temel, H., Aydoğan, A. & Tanyolaç, M. B. Major quantitative trait loci for flowering time in lentil. *Turk. J. Agric. For.* **39**, 588–595 (2015).
29. Daba, K., Deokar, A., Banniza, S., Warkentin, T. D. & Tar'an, B. QTL mapping of early flowering and resistance to ascochyta blight in chickpea. *Genome* **59**, 413–425 (2016).
30. Mallikarjuna, B. P. *et al.* Molecular mapping of flowering time major genes and QTLs in chickpea (*Cicer arietinum* L.). *Front. Plant Sci.* **8**, 1140 (2017).
31. Kumar, J., Gupta, S., Gupta, D. S. & Singh, N. P. Identification of QTLs for agronomic traits using association mapping in lentil. *Euphytica* **214**, 75 (2018).
32. Ridge, S. *et al.* The chickpea early flowering 1 (*efl1*) locus is an ortholog of Arabidopsis *ELF3*. *Plant Physiol.* **175**, 802–815 (2017).
33. Cruz-Izquierdo, S. *et al.* Comparative genomics to bridge *Vicia faba* with model and closely-related legume species: Stability of QTLs for flowering and yield-related traits. *Theor. Appl. Genet.* **125**, 1767–1782 (2012).
34. Satovic, Z. *et al.* A reference consensus genetic map for molecular markers and economically important traits in faba bean (*Vicia faba* L.). *BMC Genom.* **14**, 932 (2013).
35. Sallam, A., Dhanapal, A. P. & Liu, S. Association mapping of winter hardiness and yield traits in faba bean (*Vicia faba* L.). *Crop Pasture Sci.* **67**, 55 (2016).
36. Catt, S. C., Braich, S., Kaur, S. & Paull, J. G. QTL detection for flowering time in faba bean and the responses to ambient temperature and photoperiod. *Euphytica* **213**, 125 (2017).

37. Aguilar-Benitez, D., Casimiro-Soriguer, I. & Torres, A. M. First approach to pod dehiscence in faba bean: Genetic and histological analyses. *Sci. Rep.* **10**, 17678 (2020).
38. Webb, A. *et al.* A SNP-based consensus genetic map for synteny-based trait targeting in faba bean (*Vicia faba* L.). *Plant Biotechnol. J.* **14**, 177–185 (2016).
39. Patrick, J. W. & Stoddard, F. L. Physiology of flowering and grain filling in faba bean. *Field Crops Res.* **115**, 234–242 (2010).
40. Andrés, F. & Coupland, G. The genetic basis of flowering responses to seasonal cues. *Nat. Rev. Genet.* **13**, 627–639 (2012).
41. Chan, C. S., Peng, H.-P. & Shih, M.-C. Mutations affecting light regulation of nuclear genes encoding chloroplast glyceraldehyde-3-phosphate dehydrogenase in *Arabidopsis*. *Plant Physiol.* **130**, 1476–1486 (2002).
42. Marri, L., Sparla, F., Pupillo, P. & Trost, P. Co-ordinated gene expression of photosynthetic glyceraldehyde-3-phosphate dehydrogenase, phosphoribulokinase, and CP12 in *Arabidopsis thaliana*. *J. Exp. Bot.* **56**, 73–80 (2005).
43. Pierre, J.-B., Huguet, T., Barre, P., Huyghe, C. & Julier, B. Detection of QTLs for flowering date in three mapping populations of the model legume species *Medicago truncatula*. *Theor. Appl. Genet.* **117**, 609–620 (2008).
44. Xia, R. *et al.* ROR1/RPA2A, a putative replication protein A2, functions in epigenetic gene silencing and in regulation of meristem development in *Arabidopsis*. *Plant Cell* **18**, 85–103 (2006).
45. Sheldon, C. C. *et al.* The FLF MADS box gene: A repressor of flowering in *Arabidopsis* regulated by vernalization and methylation. *Plant Cell* **11**, 445–458 (1999).
46. Michaels, S. D. & Amasino, R. M. *FLOWERING LOCUS C* encodes a novel MADS domain protein that acts as a repressor of flowering. *Plant Cell* **11**, 949–956 (1999).
47. Bastow, R. *et al.* Vernalization requires epigenetic silencing of *FLC* by histone methylation. *Nature* **427**, 164–167 (2004).
48. Boex-Fontvieille, E. *et al.* Photosynthetic control of *Arabidopsis* leaf cytoplasmic translation initiation by protein phosphorylation. *PLoS ONE* **8**, e70692 (2013).
49. Bush, M. S. *et al.* eIF4A RNA helicase associates with cyclin-dependent protein kinase A in proliferating cells and is modulated by phosphorylation. *Plant Physiol.* **172**, 128–140 (2016).
50. Lellis, A. D. *et al.* Deletion of the eIFiso4G subunit of the *Arabidopsis* eIFiso4F translation initiation complex impairs health and viability. *Plant Mol. Biol.* **74**, 249–263 (2010).
51. Durand, E. *et al.* Flowering time in maize: Linkage and epistasis at a major effect locus. *Genetics* **190**, 1547–1562 (2012).
52. Iglesias, F. M. *et al.* Correction: The *Arabidopsis* DNA polymerase δ has a role in the deposition of transcriptionally active epigenetic marks, development and flowering. *PLoS Genet.* **11**, e1005318 (2015).
53. del Olmo, I. *et al.* *EARLY IN SHORT DAYS 7 (ESD7)* encodes the catalytic subunit of DNA polymerase epsilon and is required for flowering repression through a mechanism involving epigenetic gene silencing. *Plant J.* **61**, 623–636 (2010).
54. Wang, X. J., Cao, X. L. & Hong, Y. Isolation and characterization of flower-specific transcripts in *Acacia mangium*. *Tree Physiol.* **25**, 167–178 (2005).
55. Liu, S. *et al.* Genetic analysis and fine mapping of *LH1* and *LH2*, a set of complementary genes controlling late heading in rice (*Oryza sativa* L.). *Breed. Sci.* **62**, 310–319 (2012).
56. Klose, R. J., Kallin, E. M. & Zhang, Y. JmjC-domain-containing proteins and histone demethylation. *Nat. Rev. Genet.* **7**, 715–727 (2006).
57. Kouzarides, T. Chromatin modifications and their function. *Cell* **128**, 693–705 (2007).
58. Han, Y. *et al.* Genome-wide analysis of soybean JmjC domain-containing proteins suggests evolutionary conservation following whole-genome duplication. *Front. Plant Sci.* **7**, 1800 (2016).
59. Dutta, A., Choudhary, P., Caruana, J. & Raina, R. JMJ27, an *Arabidopsis* H3K9 histone demethylase, modulates defense against *Pseudomonas syringae* and flowering time. *Plant J.* **91**, 1015–1028 (2017).
60. Zheng, S. *et al.* The *Arabidopsis* H3K27me3 demethylase JUMONJI 13 is a temperature and photoperiod dependent flowering repressor. *Nat. Commun.* **10**, 1303 (2019).
61. Yaish, M. W., Colasanti, J. & Rothstein, S. J. The role of epigenetic processes in controlling flowering time in plants exposed to stress. *J. Exp. Bot.* **62**, 3727–3735 (2011).
62. Cubero, J. I. & Suso, M.-J. Primitive and modern forms of *Vicia faba*. *Die Kulturpflanze* **29**, 137–145 (1981).
63. Patto, M. C. V., Torres, A. M., Koblikova, A., Macas, J. & Cubero, J. I. Development of a genetic composite map of *Vicia faba* using F2 populations derived from trisomic plants. *Theor. Appl. Genet.* **98**, 736–743 (1999).
64. Román, B. *et al.* Development of a composite map in *Vicia faba*, breeding applications and future prospects. *Theor. Appl. Genet.* **108**, 1079–1088 (2004).
65. Ellwood, S. R. *et al.* Construction of a comparative genetic map in faba bean (*Vicia faba* L.); conservation of genome structure with *Lens culinaris*. *BMC Genom.* **9**, 380 (2008).
66. R Core Team. *R: A Language and Environment for Statistical Computing* (R Foundation for Statistical Computing, 2020).
67. Pflieger, S., Lefebvre, V. & Causse, M. The candidate gene approach in plant genetics: A review. *Mol. Breed.* <https://doi.org/10.1023/a:1011605013259> (2001).
68. Ocaña, S. *et al.* Large-scale transcriptome analysis in faba bean (*Vicia faba* L.) under *Ascochyta fabae* infection. *PLoS ONE* **10**, e0135143 (2015).
69. Torres, A. M., Weeden, N. F. & Martín, A. Linkage among isozyme, RFLP and RAPD markers in *Vicia faba*. *Theor. Appl. Genet.* **85**, 937–945 (1993).
70. Medrano, R. F. V. & de Oliveira, C. A. Guidelines for the tetra-primer ARMS-PCR technique development. *Mol. Biotechnol.* **56**, 599–608 (2014).
71. Van Ooijen, J. W. *JoinMap* 4, Software for the Calculation of Genetic Linkage Maps in Experimental Populations* Vol. 33, 4 (Kyazma BV, 2006).
72. Kosambi, D. D. The estimation of map distances from recombination values. *Ann. Eugen.* **12**, 172–175 (1943).
73. van Ooijen, J. W. *MapQTL* 5 Software for the Mapping of Quantitative Trait Loci in Experimental Populations* (Kyazma B.V., 2004).
74. Lander, E. S. & Botstein, D. Mapping mendelian factors underlying quantitative traits using RFLP linkage maps. *Genetics* **121**, 185–199 (1989).
75. van Ooijen, J. W. Accuracy of mapping quantitative trait loci in autogamous species. *Theor. Appl. Genet.* **84**, 803–811 (1992).
76. Jansen, R. C. Interval mapping of multiple quantitative trait loci. *Genetics* **135**, 205–211 (1993).
77. Jansen, R. C. Controlling the type I and type II errors in mapping quantitative trait loci. *Genetics* **138**, 871–881 (1994).
78. Jansen, R. C. & Stam, P. High resolution of quantitative traits into multiple loci via interval mapping. *Genetics* **136**, 1447–1455 (1994).
79. Churchill, G. A. & Doerge, R. W. Empirical threshold values for quantitative trait mapping. *Genetics* **138**, 963–971 (1994).
80. Voorrips, R. E. MapChart: Software for the graphical presentation of linkage maps and QTLs. *J. Hered.* **93**, 77–78 (2002).

Acknowledgements

This research was supported by funding from projects RTA2017-00041 co-financed by ERDF and PP.AVA. AVA2019.030 by the Junta de Andalucía. DAB acknowledges his Ph.D. fellowship INIA-CCAA. The authors are grateful to Dr. A. Di Pietro for critical reading of the manuscript and Dr. S. Cruz-Izquierdo and Dr. F. Maalouf for the phenotypic data collection.

Author contributions

D.A.-B., I.C.-S. and A.M.T. conceived and designed the study and laboratory experiments. F.M. performed the flowering evaluation in Terbol, Lebanon. D.A.-B. and I.C.-S. performed the genotyping and mapping studies and analyzed the data. D.A.-B., I.C.-S. and A.M.T. interpreted the results and wrote the manuscript.

Competing interests

The authors declare no competing interests.

Additional information

Supplementary Information The online version contains supplementary material available at <https://doi.org/10.1038/s41598-021-92680-4>.

Correspondence and requests for materials should be addressed to A.M.T.

Reprints and permissions information is available at www.nature.com/reprints.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2021